



VANDERBILT UNIVERSITY

# Effects of Cholesterol Enhancement on the Cell Properties in the Presence of Graphene

VANDERBILT UNIVERSITY  
MEDICAL CENTER

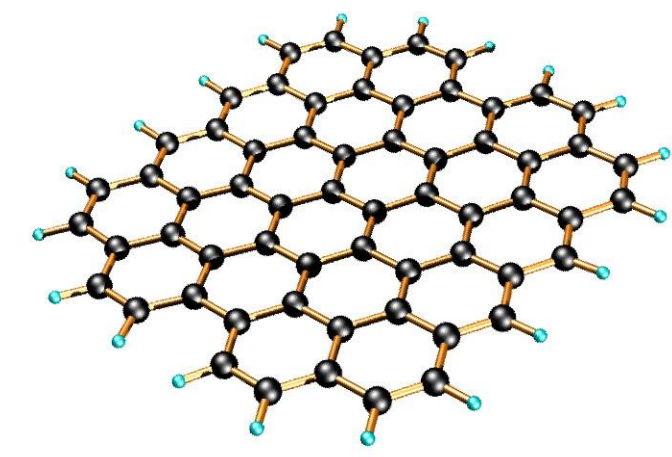
Jason A. Ray-Alfaro<sup>1</sup>, Kristina E. Kitko<sup>1,2</sup>, Roman Lazarenko<sup>1</sup>, Tu Hong<sup>3</sup>, Da Ying<sup>3</sup>, Ya-Qiong Xu<sup>3</sup>, Qi Zhang<sup>1</sup>

<sup>1</sup>Department of Pharmacology, Vanderbilt University, Nashville TN 37240

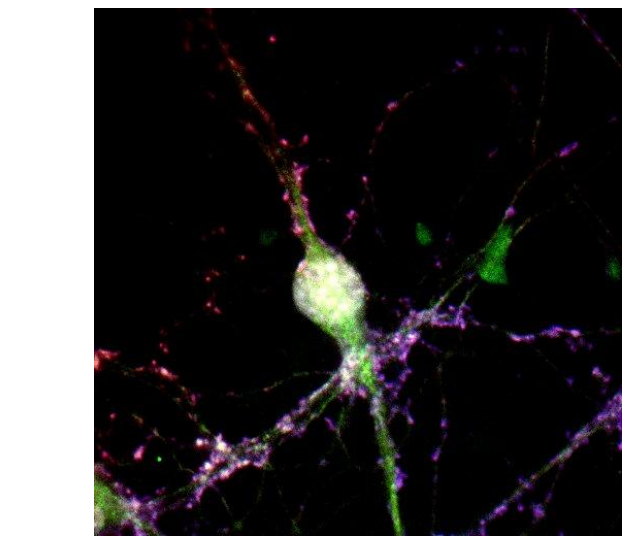
<sup>2</sup>Program in Interdisciplinary Materials Science, Vanderbilt University, Nashville TN 37235

<sup>3</sup>Department of Electrical and Computer Engineering, Vanderbilt University, Nashville, TN 37235

## Introduction



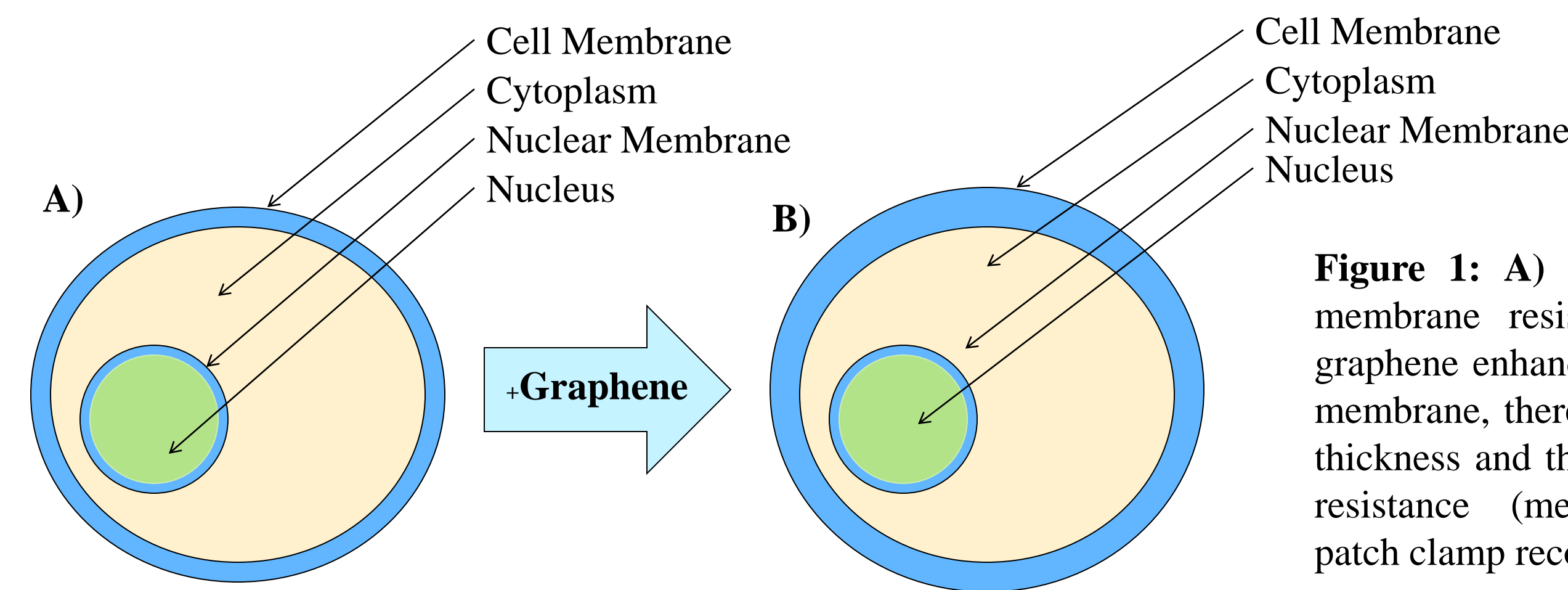
**Motivation:** Graphene is a two-dimensional carbon crystal with remarkable mechanical strength and a singular electrical conductivity, a combination that has led to interest in biomedical application. However, this demands a clear understanding of graphene's interaction with cell surface molecules and its impact on cell function.



### Objectives:

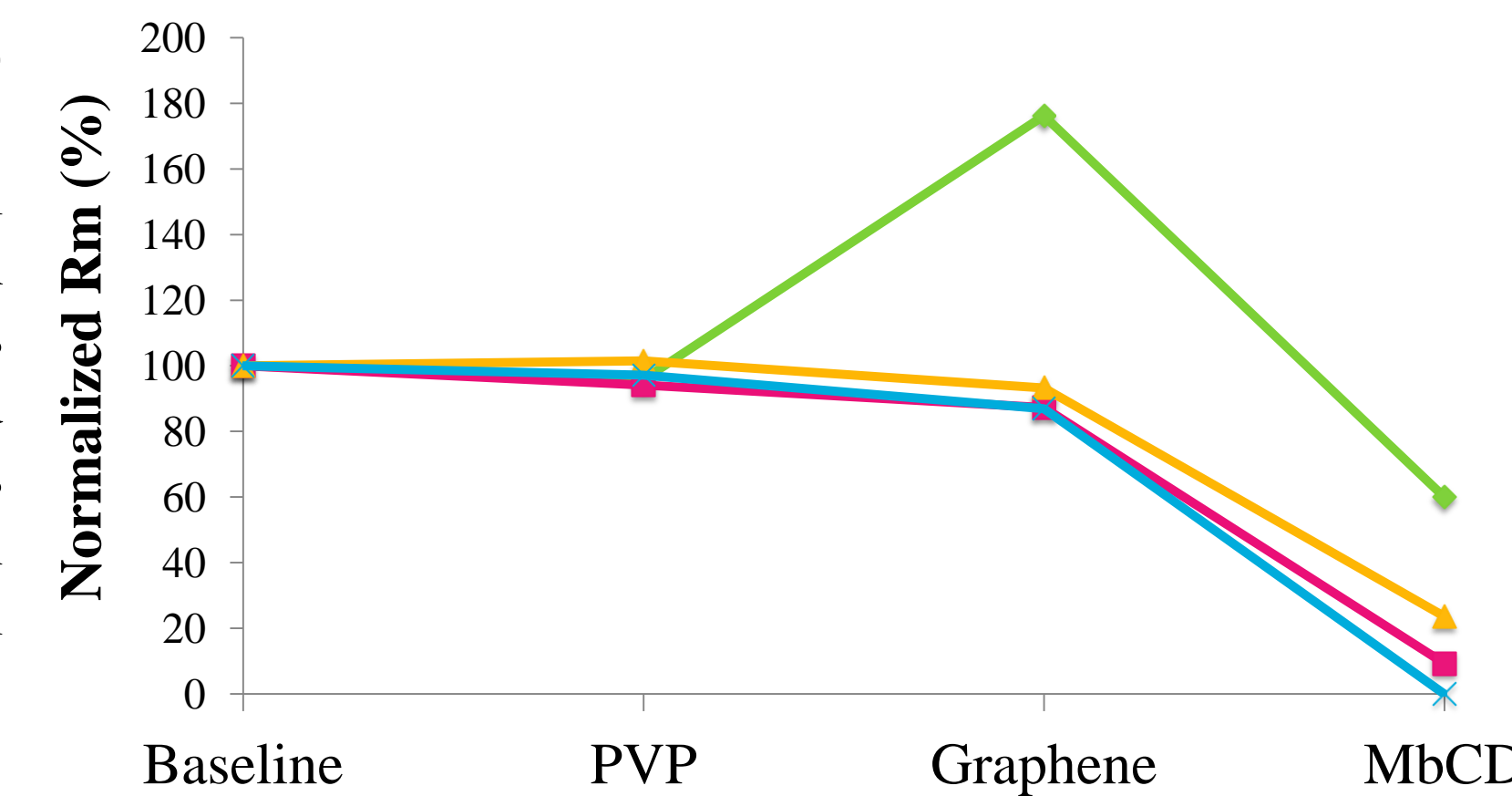
- To analyze the cholesterol concentration of culture media in the presence of graphene using spectral analysis
- To modify cell properties using graphene and demonstrate a relationship between graphene concentration and membrane resistance
- To highlight the importance of proper procedure when producing cell cultures to prevent cortex contamination

## Graphene may Increase Membrane Resistance



**Figure 1:** A) Untreated cell, normal membrane resistance. B) Addition of graphene enhances the cholesterol in the membrane, thereby increasing its overall thickness and thus whole cell membrane resistance (measured via whole-cell patch clamp recording).

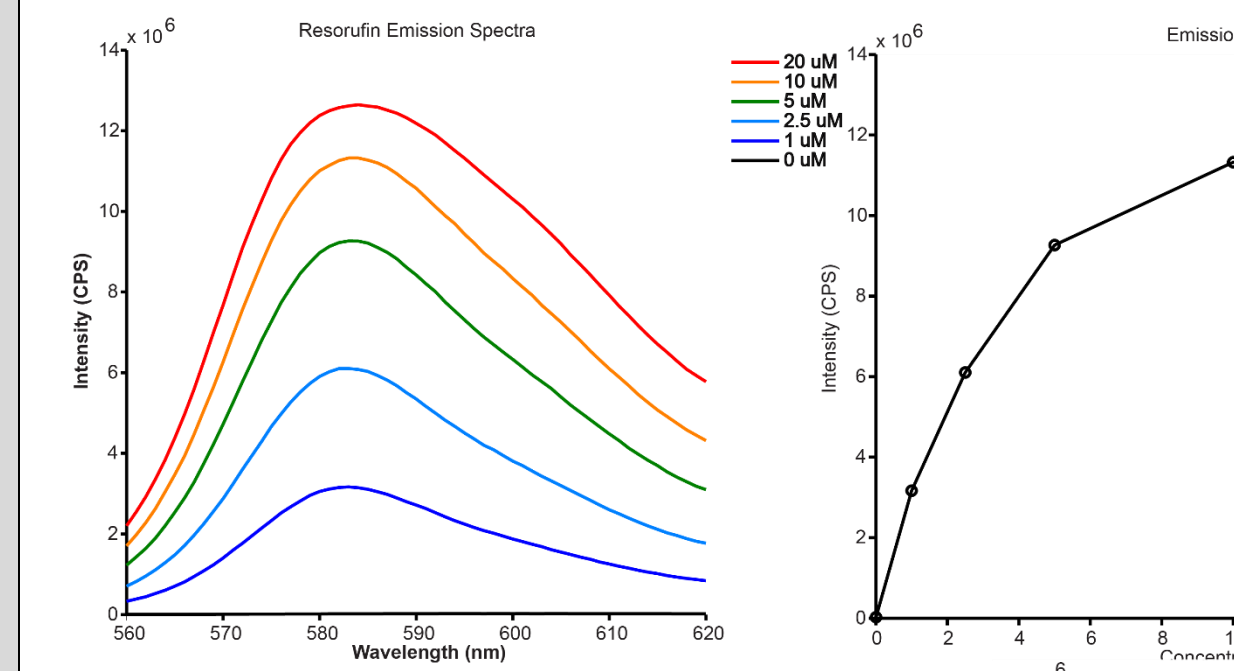
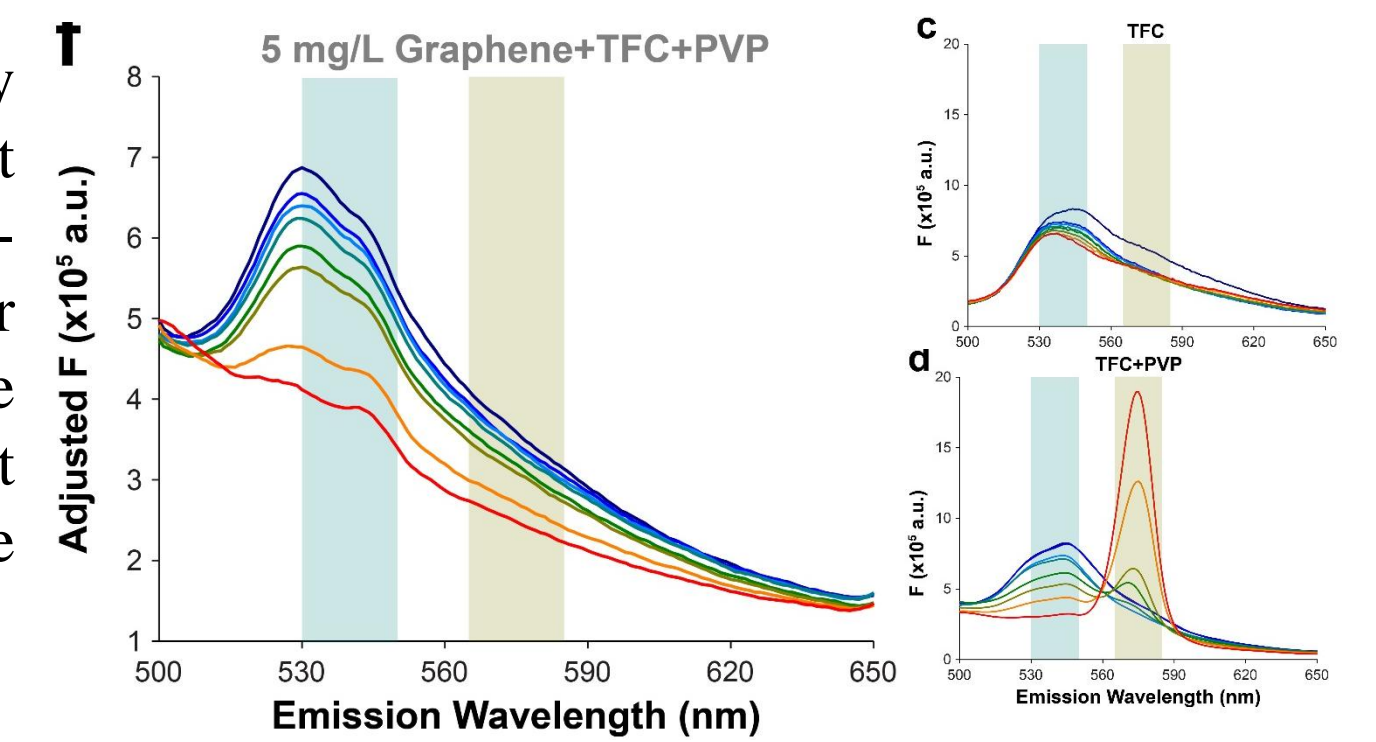
### Membrane Resistance



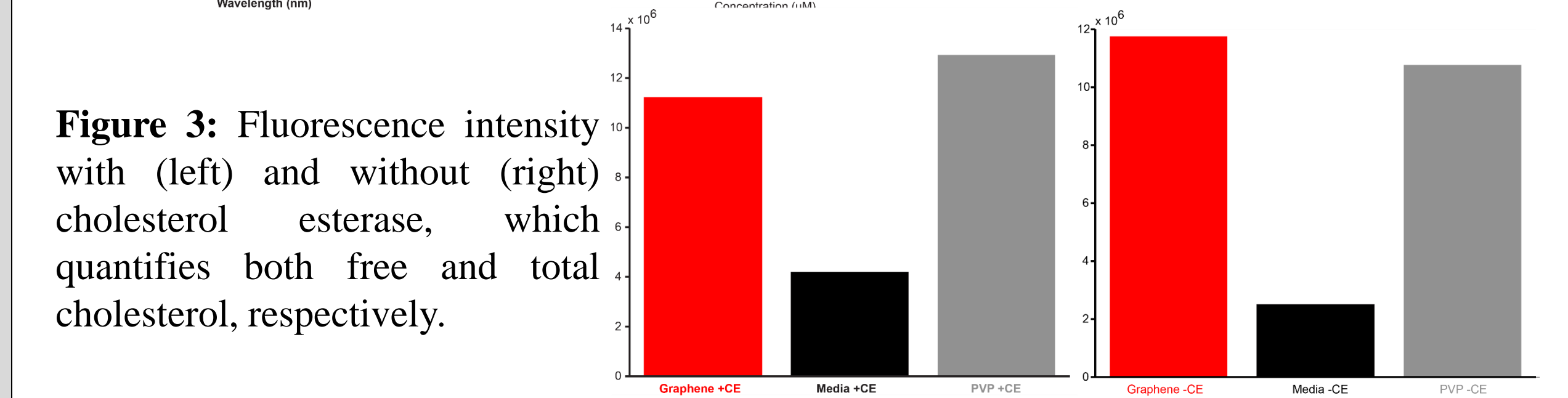
**Figure 2:** Data from four records of membrane resistance ( $R_m$ ) measurement. Cells were treated in 4 different conditions: Baseline, Polyvinylpyrrolidone (PVP) solution, 40 $\mu$ m/ml Graphene in PVP solution, and Methyl-beta-cyclodextrin (M $\beta$ CD).  $R_m$  is normalized with the baseline value of each cell as 100%. PVP solution is a sham control as graphene was dissolved in PVP solution. M $\beta$ CD deprives membrane cholesterol and thus reduces  $R_m$ . One out of four cells showed increased  $R_m$  upon graphene application, highlighting the importance of additional recordings.

## Testing the Graphene and Cholesterol Interaction

**Figure 1:** Spectrofluorometry measurements show the time-dependent fluorescence quenching of a graphene-fluorophore solution, unique from either the fluorophore alone (right top) or the fluorophore and the solubilizer (right bottom), suggestive of a graphene interaction.



**Figure 2:** Resorufin emission spectra calibration. Resorufin is an end product of the cholesterol reaction, thus used as a baseline to verify assay functionality.

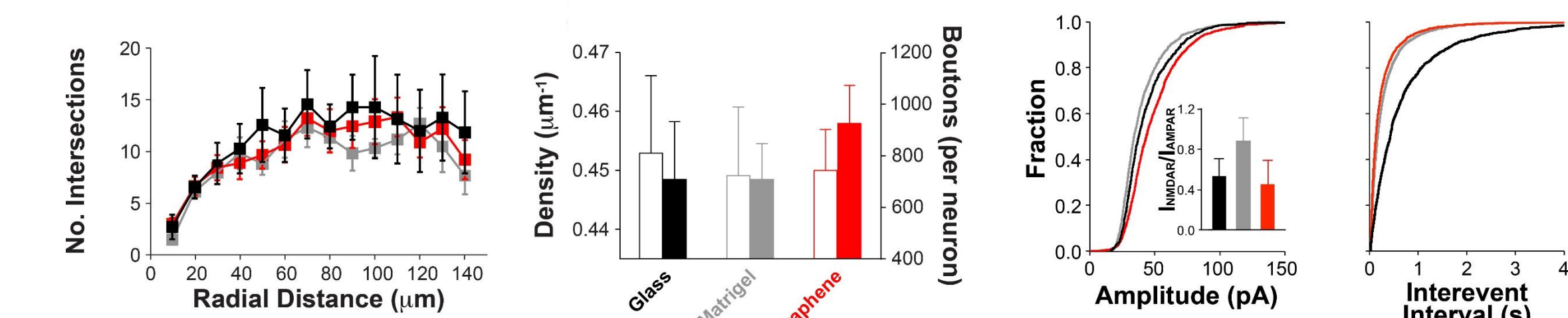


**Figure 3:** Fluorescence intensity with (left) and without (right) cholesterol esterase, which quantifies both free and total cholesterol, respectively.

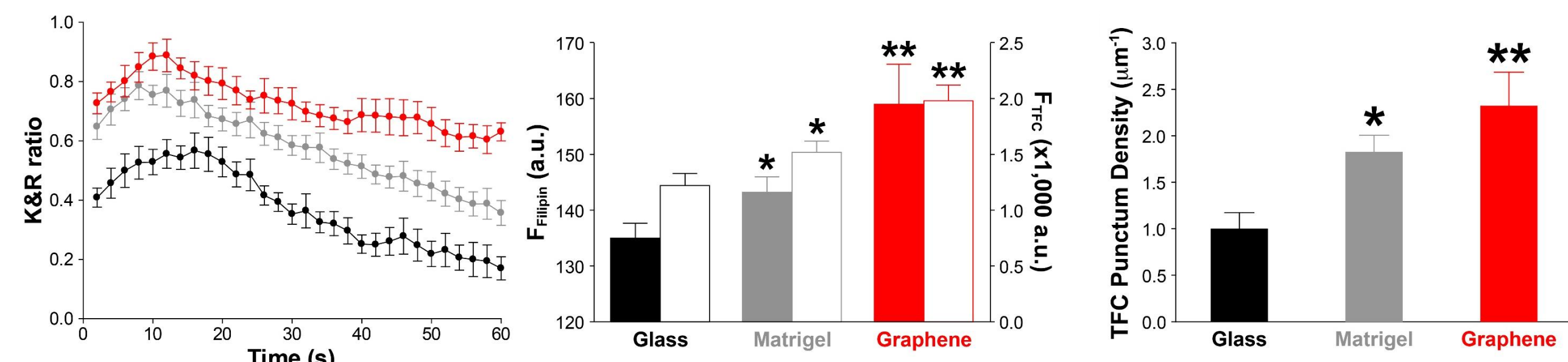
## Previous Research

Previous research concluded:

- Graphene does not cause significant changes in neuronal morphology



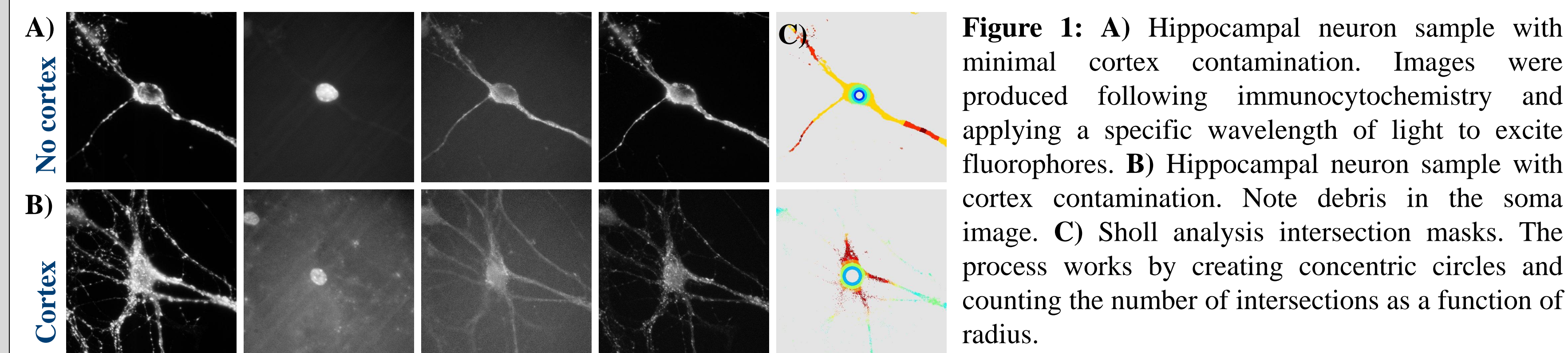
- An electrophysiological change in frequency, but not amplitude of mEPSCs suggests that most likely the impact is presynaptic



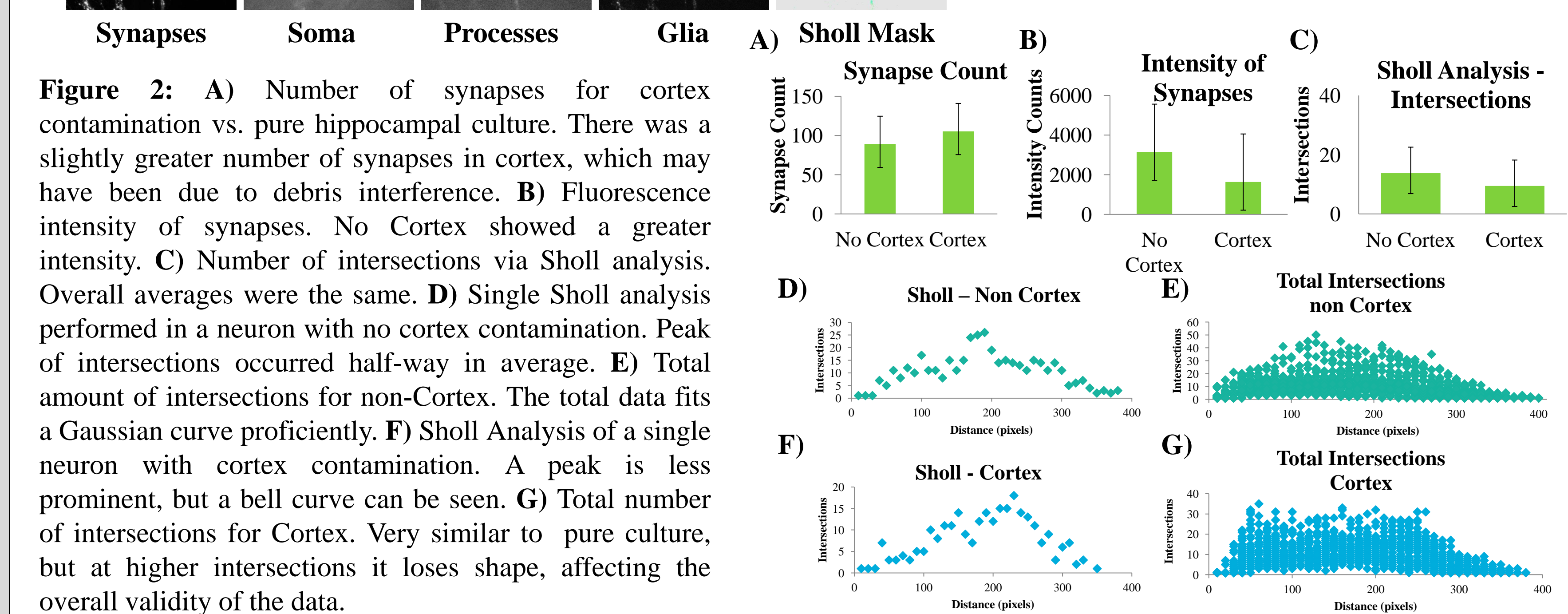
- Synaptic vesicle tracking combined with cholesterol imaging implicates that graphene increases both membrane cholesterol content and membrane curvature

- As such changes are limited and defined, bare graphene may be used as both a growth substrate and a field effect transistor to detect membrane potential changes

## Impact of Cortex on Hippocampal Networks



**Figure 1:** A) Hippocampal neuron sample with minimal cortex contamination. Images were produced following immunocytochemistry and applying a specific wavelength of light to excite fluorophores. B) Hippocampal neuron sample with cortex contamination. Note debris in the soma image. C) Sholl analysis intersection masks. The process works by creating concentric circles and counting the number of intersections as a function of radius.



**Figure 2:** A) Number of synapses for cortex contamination vs. pure hippocampal culture. There was a slightly greater number of synapses in cortex, which may have been due to debris interference. B) Fluorescence intensity of synapses. No Cortex showed a greater intensity. C) Number of intersections via Sholl analysis. Overall averages were the same. D) Single Sholl analysis performed in a neuron with no cortex contamination. Peak of intersections occurred half-way in average. E) Total amount of intersections for non-Cortex. The total data fits a Gaussian curve proficiently. F) Sholl Analysis of a single neuron with cortex contamination. A peak is less prominent, but a bell curve can be seen. G) Total number of intersections for Cortex. Very similar to pure culture, but at higher intersections it loses shape, affecting the overall validity of the data.

## Results and Future Work

- Cortex has a slight negative impact on hippocampal networks. The debris increases the uncertainty of analysis done via immunocytochemistry as the antibodies will also stain the cortex
- Preliminary data shows implicates that graphene may indeed increase membrane resistance, although additional verification is necessary
- Further spectral analysis of fluorophore interaction with graphene reveals a time-dependent fluorescence decay that is unique from that of the fluorophore, consistent with an interaction involving energy transfer

**Future Work** will include additional patch clamping experiments on different cell types to further study the relationship between chronic or acute graphene exposure and membrane resistance

## Acknowledgements



Research was made possible in part through the Vanderbilt Institute of Nanoscale Science and Engineering (VINSE), Vanderbilt University Pharmacology with a special thanks to Qi Zhang, Ph.D. and Kristina Kitko, M.S.. This work was supported by the Nation Science Foundation: Research for Undergraduates, Grant NSF DMR-1263182.

### References:

Graphene Diagram: Hamilton, Christopher E. "Graphene." OpenStax CNX. RICE, 2012. Web.