



# THE DRUG-INDEPENDENT ROLES OF CARDIAC GEOMETRY AND TISSUE ANISOTROPY IN DEFIBRILLATION AND REENTRY

J.P. Wikswo, Ph.D. and S. F. Lin, Ph.D.

Department of Physics and Astronomy, Vanderbilt  
University, Nashville, TN 37235 USA.



# There is more to reentry and defibrillation than just channel effects ....

- Antiarrhythmic drugs primarily alter tissue excitability
- Defibrillation and reentry are also affected by tissue tissue anisotropy and cardiac geometry, which are not altered directly by drugs



# Antiarrhythmic drugs are not the entire story....

- A high percentage of patients with an implantable ICD/defibrillator also take antiarrhythmic drugs.
- Agonistic/antagonistic effects of drugs on arrhythmias, fibrillation, and defibrillatory efficacy are the subject of intense study at the microscopic level of the ion channel.
- The primary effect of antiarrhythmic drugs is to alter tissue excitability, through changes in threshold, rate of rise, and refractoriness; **antiarrhythmic drugs do not strongly affect cell-to-cell coupling.**



## The rest of the story....

- To first order, the shock-tissue interaction is determined by tissue anisotropy and cardiac geometry, and **is not altered directly by drugs.**
- The shock-tissue interaction is poorly understood: there is not yet an answer to the question “*How does defibrillation work?*”



# Possible explanations for the ability of strong stimuli to defibrillate the heart

## Discrete Mechanisms

- Intracellular junctions
- Uncoupled fiber bundles
- Patches of fat or collagen
- Bidomain heterogeneities

## Continuous Mechanisms

- Cardiac surfaces
- Fiber ends
- Tissue anisotropy
- Fiber curvature
- Fiber branching
- Strand taper
- Fiber rotation



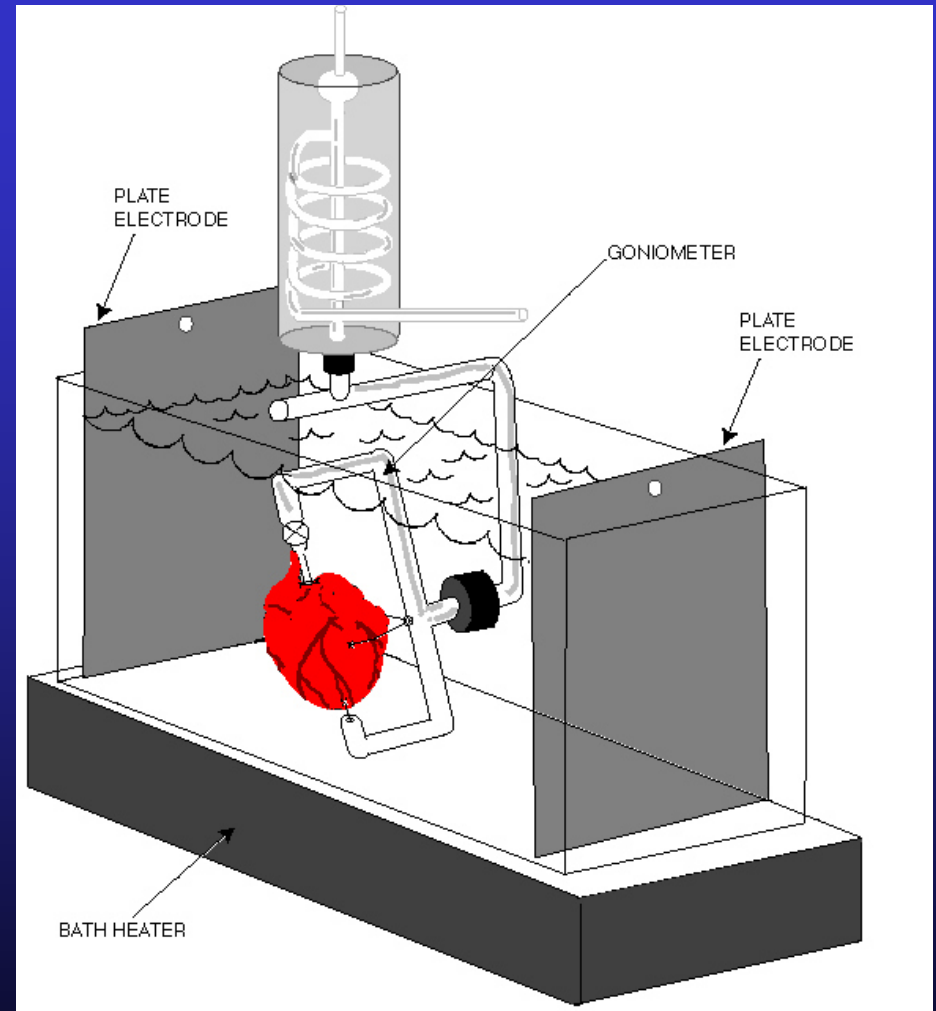
# Approach

- Optical imaging of the transmembrane potential, and the bidomain model allow us to study the shock-tissue interaction and reentry.
- We examined the rate of activation of the endocardium and epicardium during normal activation and field stimulation.

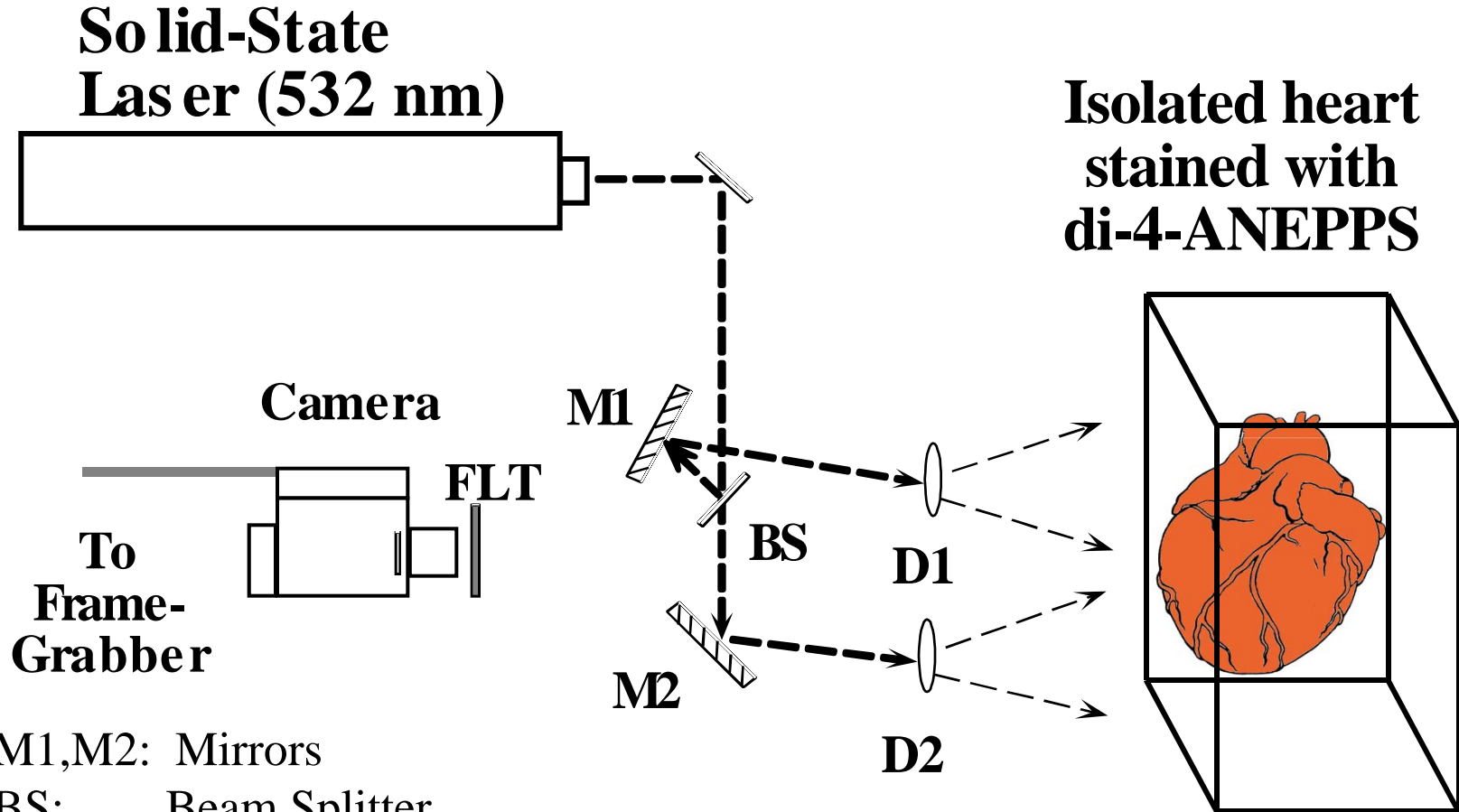


## Methods: Perfusion

- Langendorff-perfused, di-4-ANNEPPS stained, isolated rabbit hearts with atria excised (N=5)
- 10 x 10 x 15 cm<sup>3</sup> bath of Tyrode's solution at 37 °C.
- Goniometer allows 360° rotation of the axis of the heart in the imaging plane.



# Optical Imaging Setup



M1,M2: Mirrors

BS: Beam Splitter

D1, D2: Diffusers

FLT: Long-Pass Filter (590 nm)





## Methods: Imaging

- Optical imaging of  $V_m$  changes following S1 as well as during and after S2.
- High-speed (322 or 1000 frames/second) 12-bit Dalsa CCD camera.
- Heart illuminated by a diode-pumped laser with multiple, 1 mm plastic optical fibers.



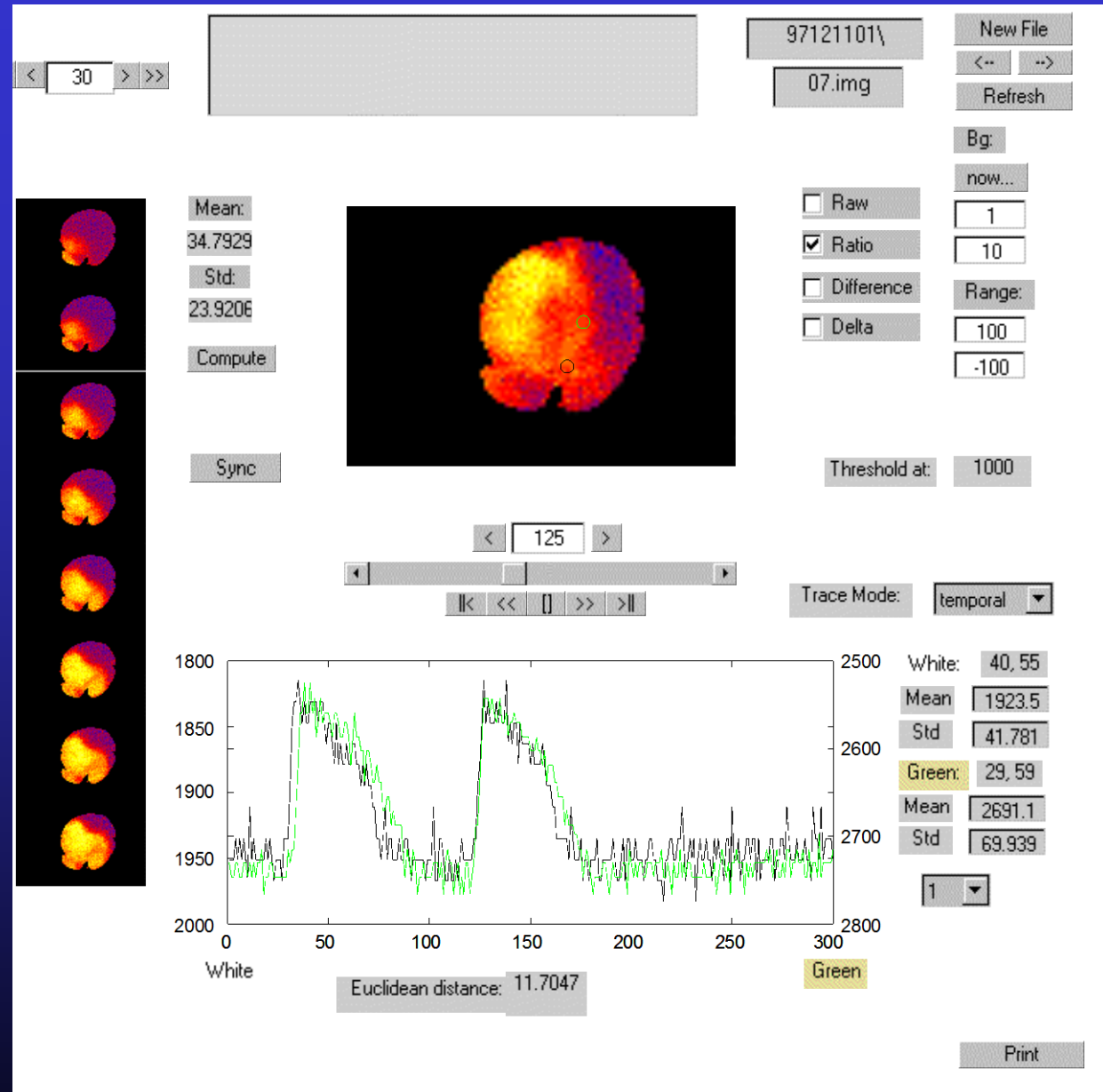
## Methods: Pacing

- 20 S1 pacing pulses to the right-ventricle 500 ms cycle length
- Custom Ventritex computer-controlled stimulator delivered a 1 ms S2 to plate electrodes at the ends of the bath to produce horizontal shock fields of up to  $\pm 33$  V/cm

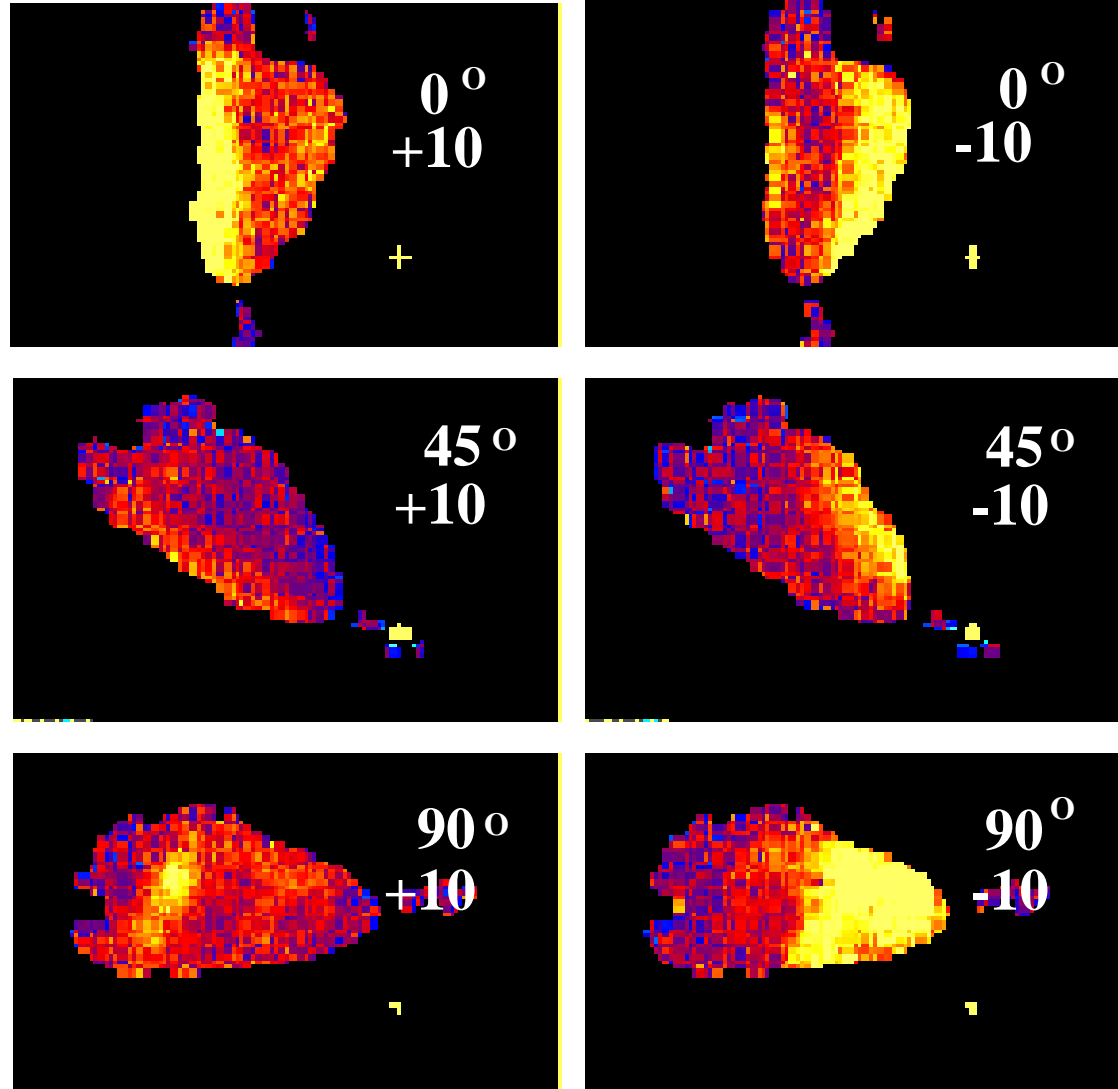


# Gus2 MatLab Data Viewing Program

Gustavo Rohde

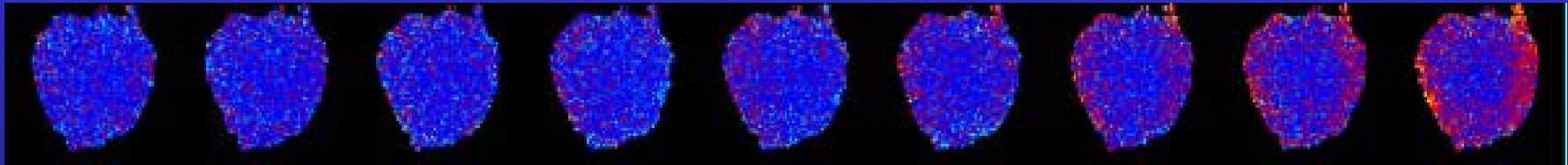


# Tissue Response to Field Shock





# Faster Response with Increasing Shock Strength



98021001\10 20 Volts/cm, 3 ms shock, 1 ms per frame



## Methods: Image Processing

- $Q(t)$  -- measure of total activation in the image

$$Q(t) = \sum_{ij} \{ \Delta F(t)/F(0) \}_{ij}, \quad i.e., \text{sum of all pixels}$$

- $p_x(t)$  -- dipole moment along the x axis

$$p_x(t) = \sum_{ij} x_i \{ \Delta F(t)/F(0) \}_{ij}, \quad i.e., \text{intensity times } x$$

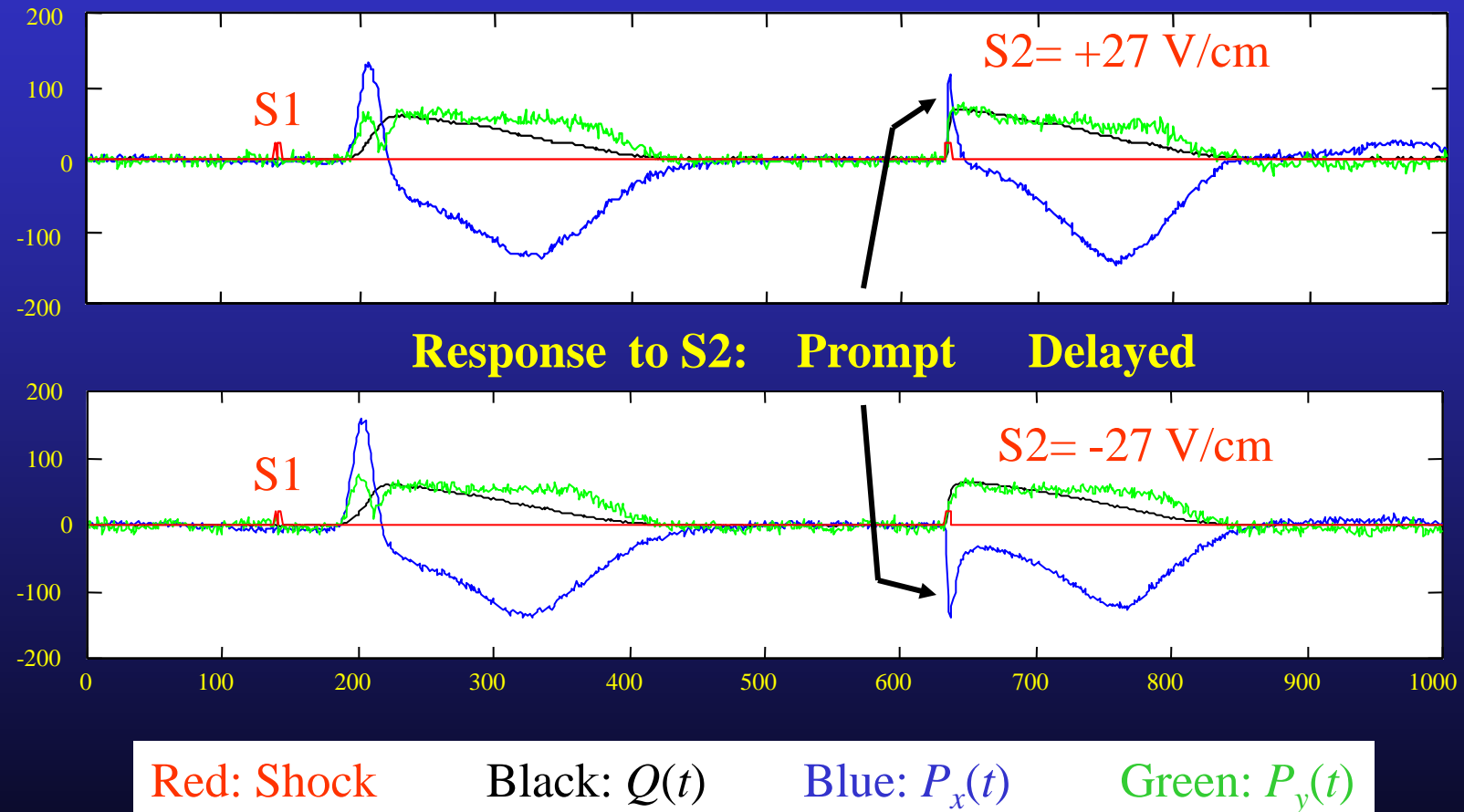
- $p_y(t)$  -- dipole moment along the y axis

$$p_y(t) = \sum_{ij} y_i \{ \Delta F(t)/F(0) \}_{ij}, \quad i.e., \text{intensity times } y$$



# $Q(t), p_x(t), p_y(t)$ for S1 and $\pm 27$ V/cm S2

- $Q(t)$  rises faster for S2 than S1
- $p_x$  prompt S2 response reverses sign
- $p_y$  prompt S2 response unchanged





# How fast is the heart being activated by S1 and S2?

- Measure maximum rate of rise

$$[dQ(t)/dt]_{max}$$

- Measure maximum value of  $Q(t)$

$$Q(t)_{max}$$

- Define activation time

$$T_{activ} = Q(t)_{max} / [dQ(t)/dt]_{max}$$



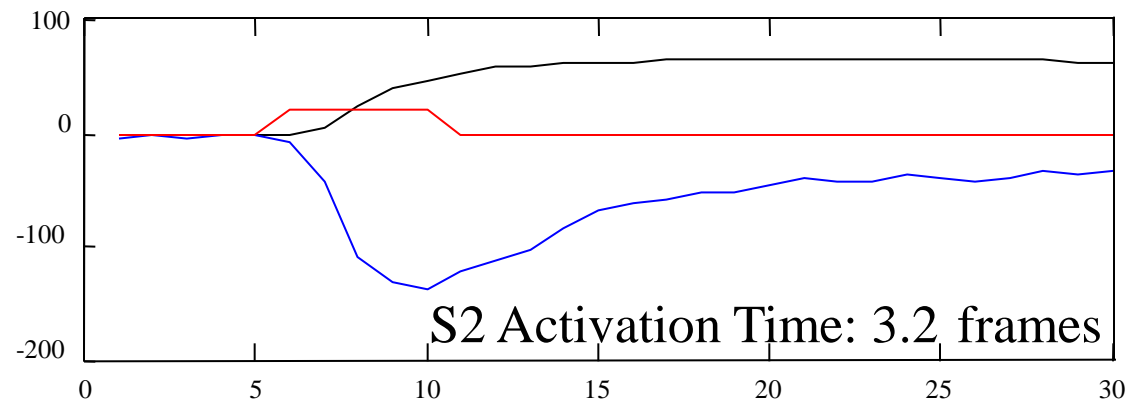
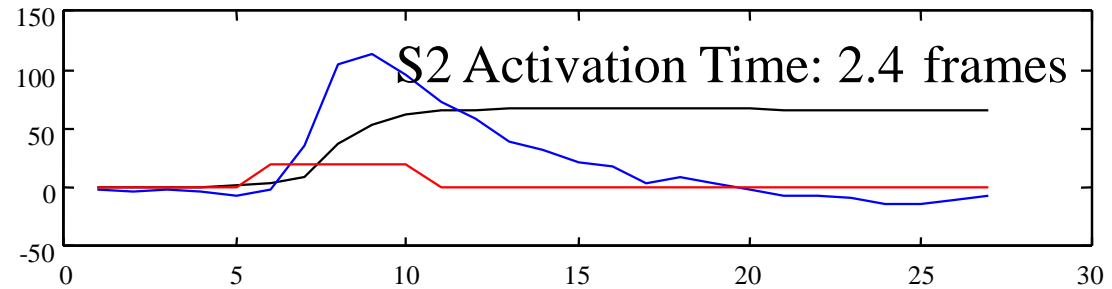
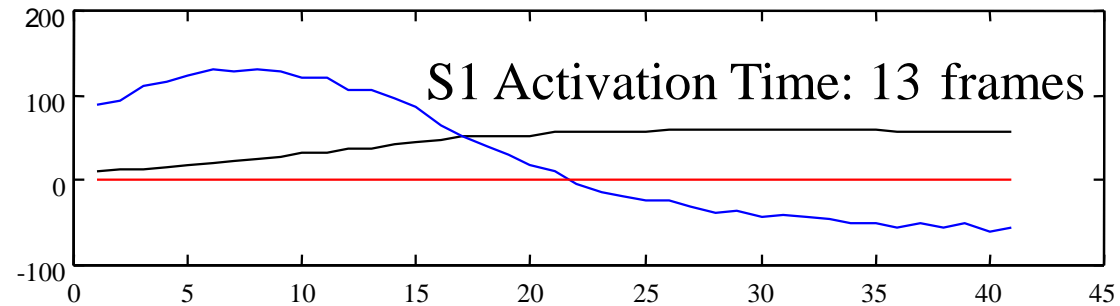


# Activation Times

S2: +27 V/m

S2: -27 V/m

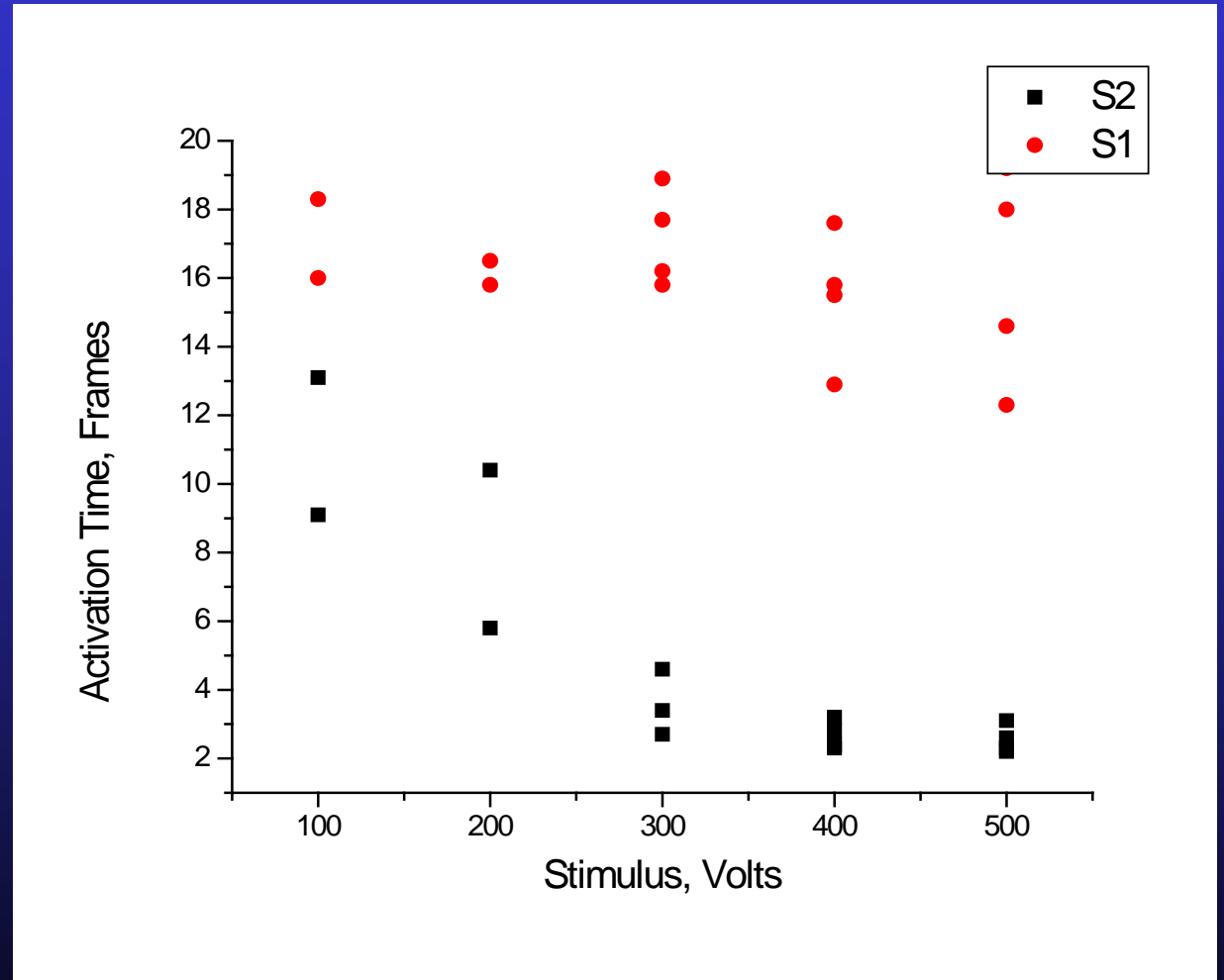
Red: Shock  
Black:  $Q(t)$   
Blue:  $P_x(t)$





# Activation Time versus S2 Field Strength

- S1 activation time independent of S2 field strength
- S2 activation time decreases with increasing S2 field strength

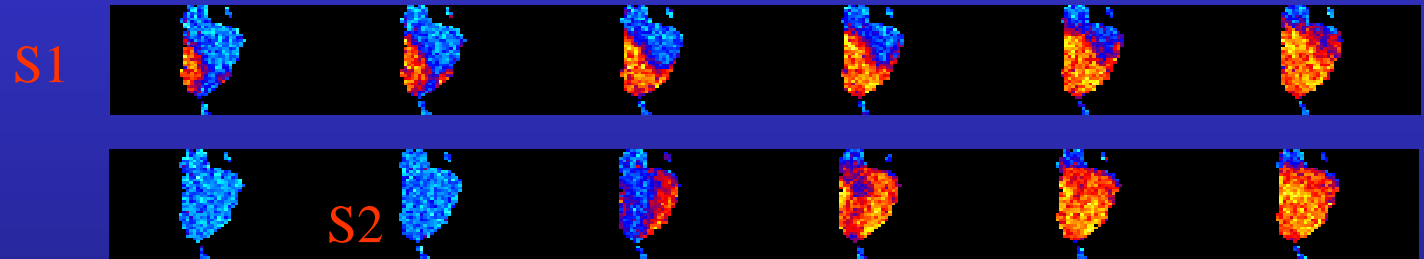




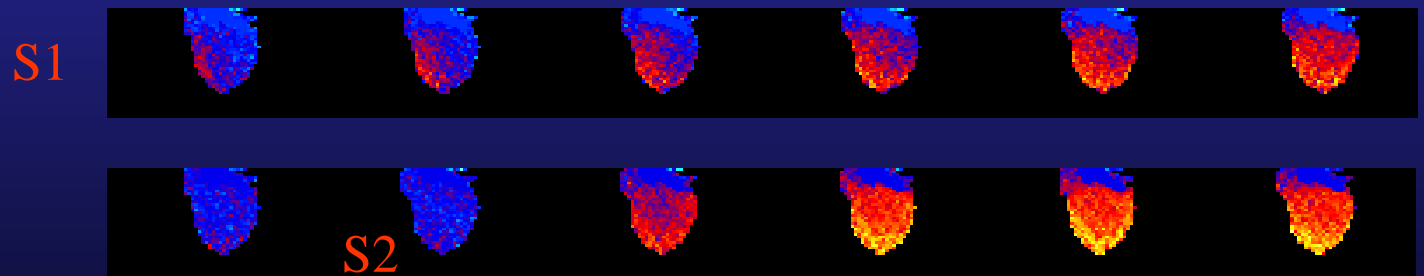
# Transmural Imaging of Endocardial Activation

- Optical fibers with tapered diffusers illuminate the endocardium
- CCD camera views activity transmurally
- No endocardial  $p_x$  dipole moment
- Rapid endocardial activation

Epicardial Images for  $S2 = +10$  V/cm



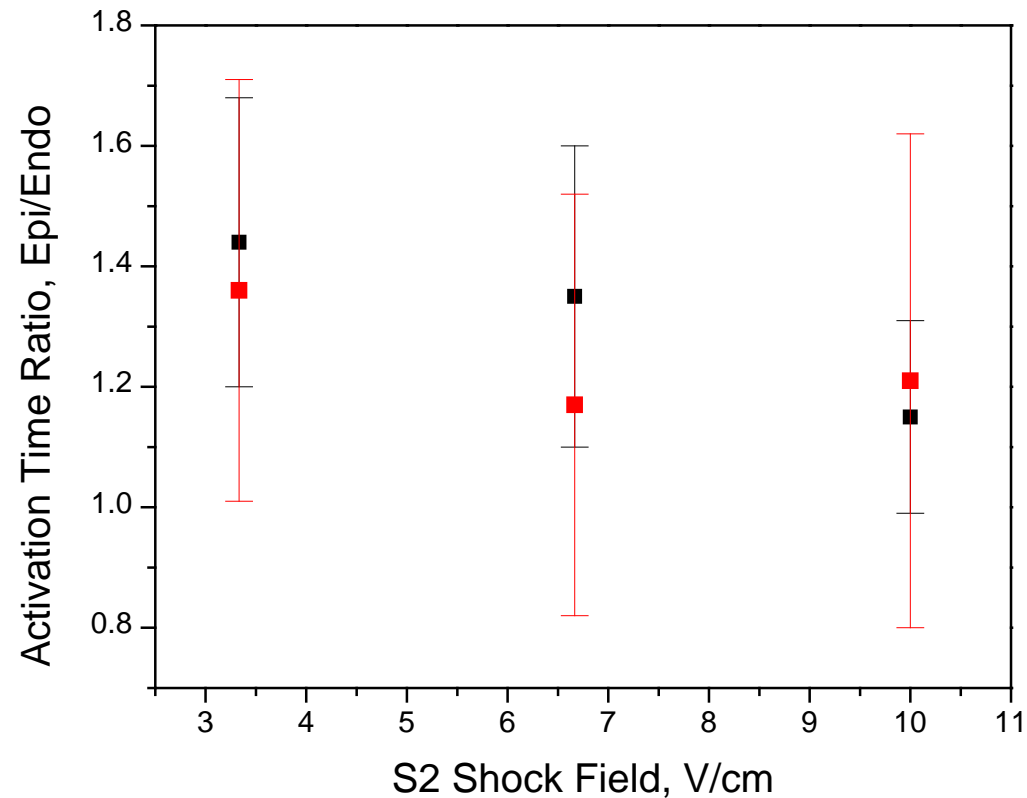
Endocardial Images for  $S2 = +10$  V/cm





# Endocardial versus Epicardial Activation Times

- Activation time decreases with increasing shock field
- Endocardial response 20%-40% faster than endocardial
- Trend: S1 response (Black) slower than S2 (Red)





# Summary

Endocardial and endocardial responses to field stimulation differ:

- Dipolar response of epicardium consistent with bidomain predictions
- Endocardium does not show dipolar response. Why?
- Endocardium activates faster than epicardial. Why?
- Not due solely to Purkinje fibers -- trabeculae and papillary muscles?

Hypothesis:

The complex geometrical structure of the endocardial surface plays an as-yet-unappreciated role in the rapid and uniform activation of the endocardium during defibrillation.