

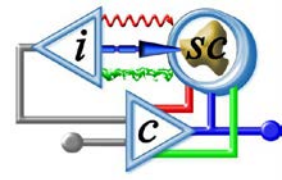
# Challenges of Single Ion-Channel Biosensors

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APS March Meeting  
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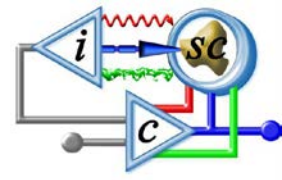
# Receptors as Biosensors



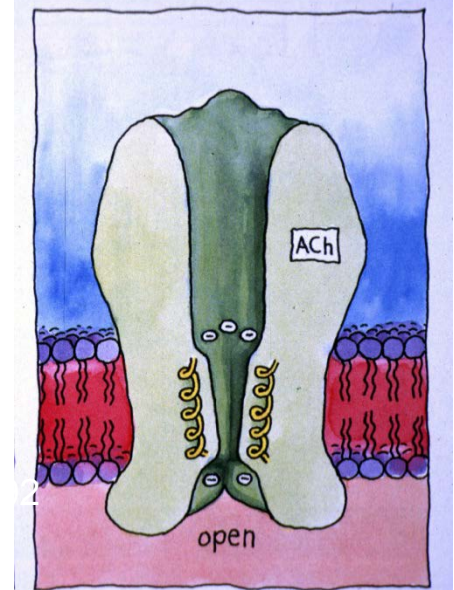
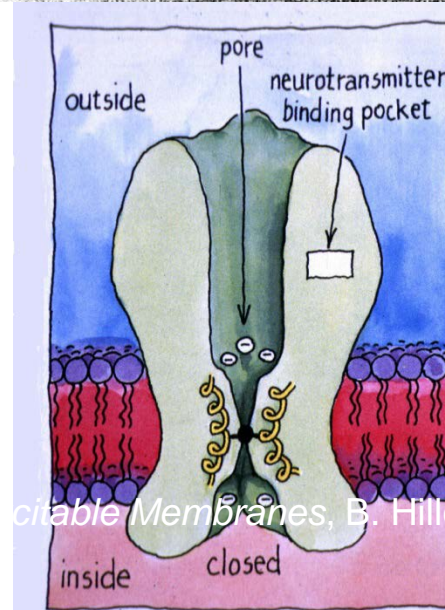
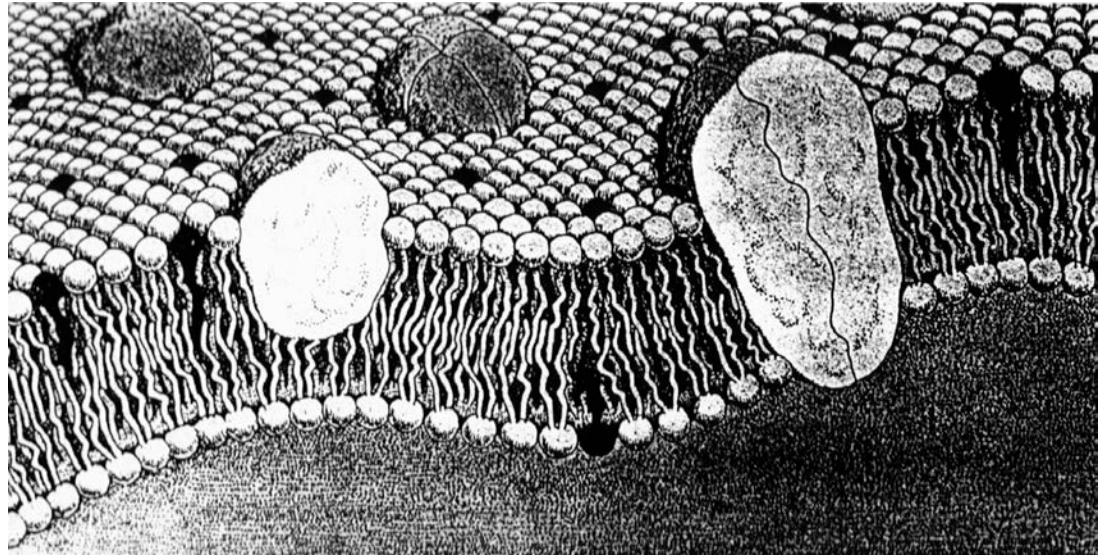
- Receptor binding can be used to detect chemical and biological agents, drugs, and environmental toxins. This can be accomplished in several ways
  - Fluorophores
  - Field effect transistors
  - Surface plasmon resonance
  - Labeled nanoparticles
  - Nanocantilevers
  - Ion Channels

# Membrane Ion Channel

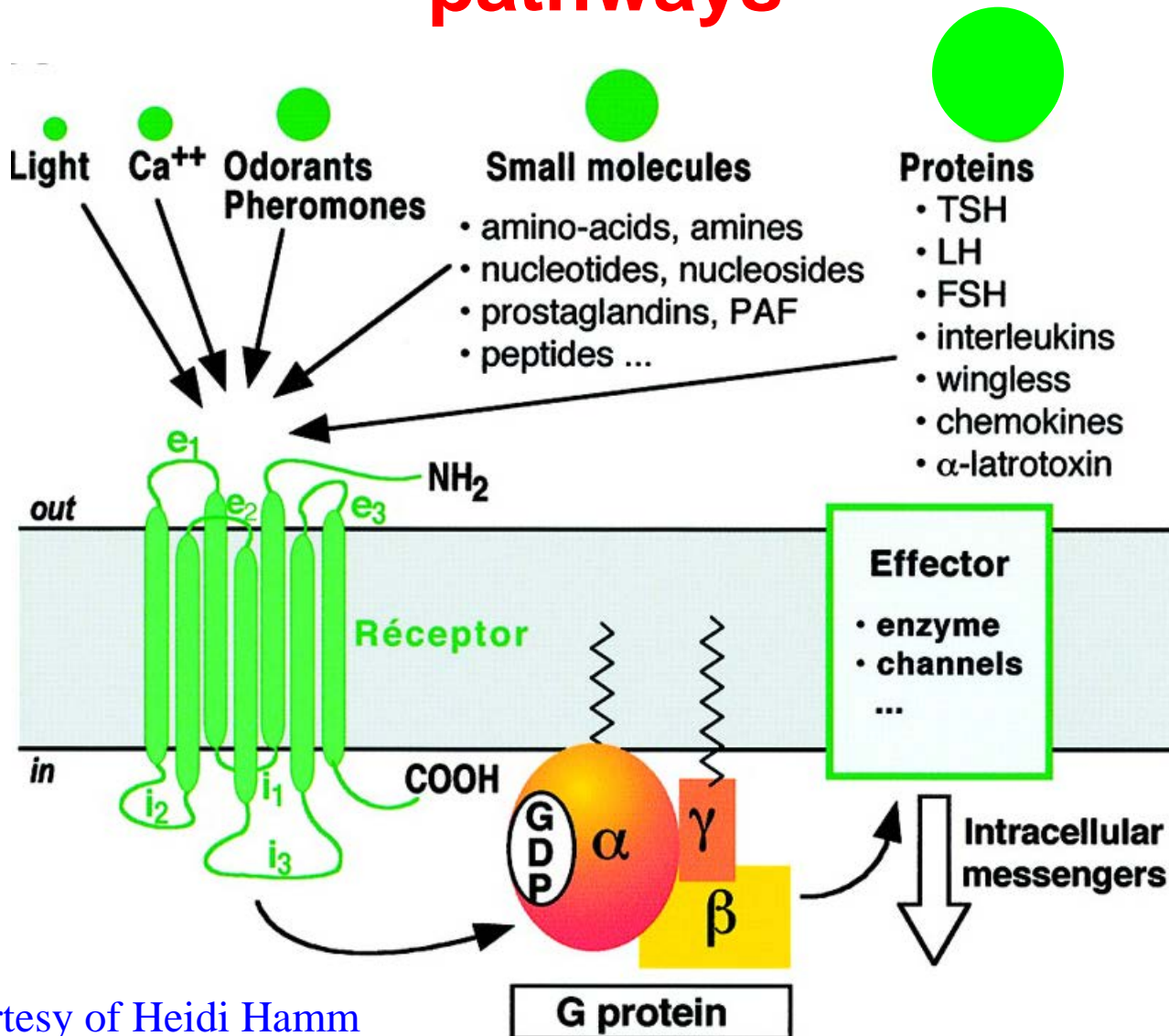
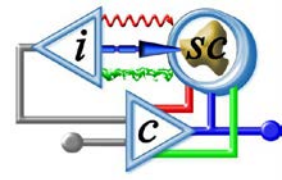
~ 1 nm pore



- Membrane-spanning protein
- MscL,  $\alpha$ HL, and connexins: large pores with no ionic selectivity.
- $K^+$  channels: ion selective and often rectification even in symmetric solutions
- Gating controlled by electric or mechanical forces and ligands
- Gated ion channels can assume discrete conducting states

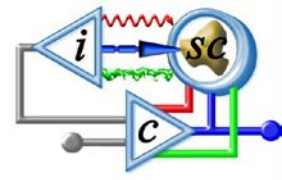


# G-protein-coupled receptors exhibit complex signalling pathways

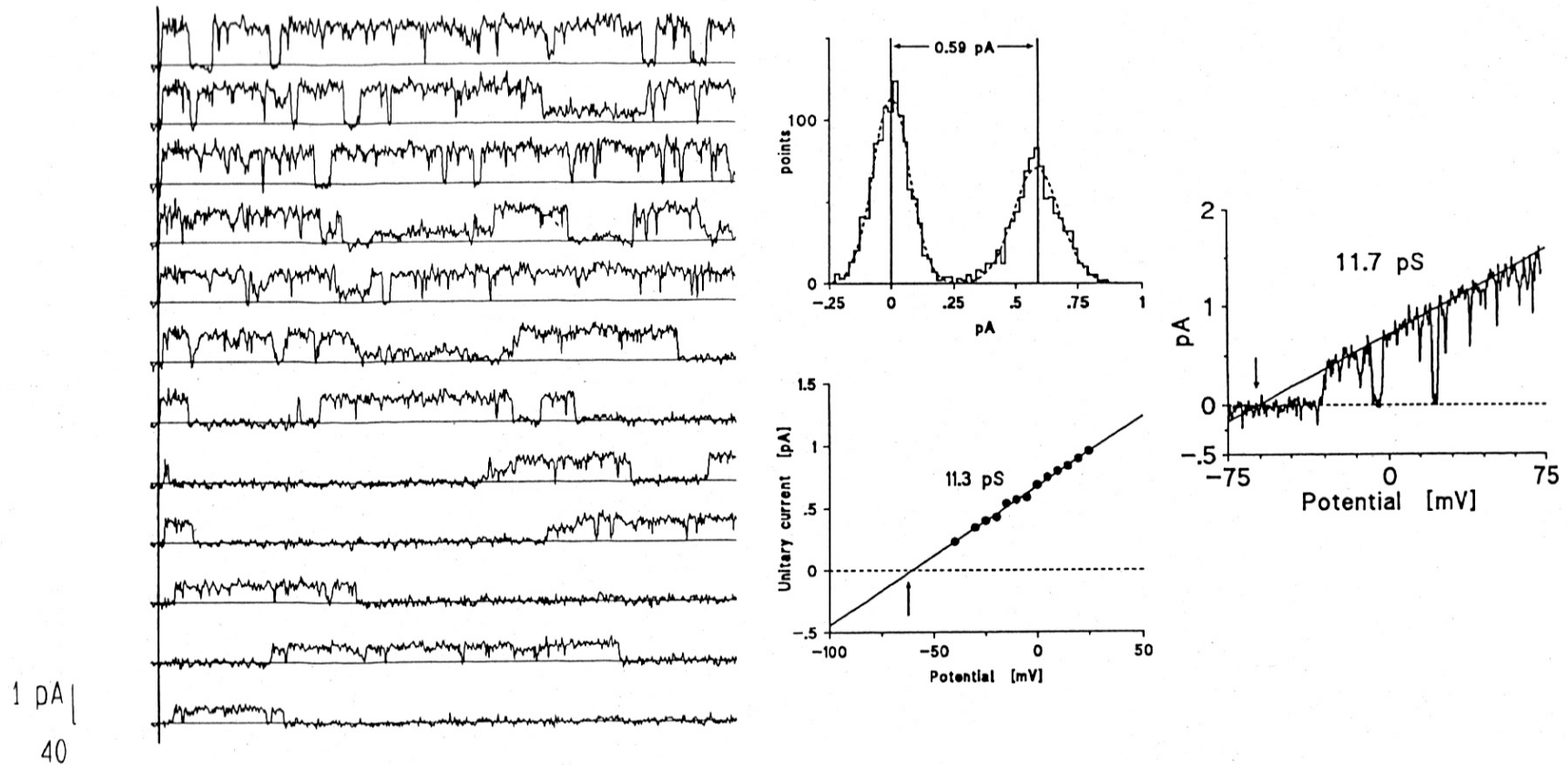


Courtesy of Heidi Hamm

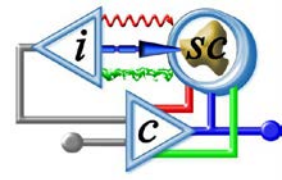




# Transmembrane ion channels have a time- and voltage- or ligand-dependent conductance that can serve as the basis for a biosensor



# A particular ligand can control channel conductance

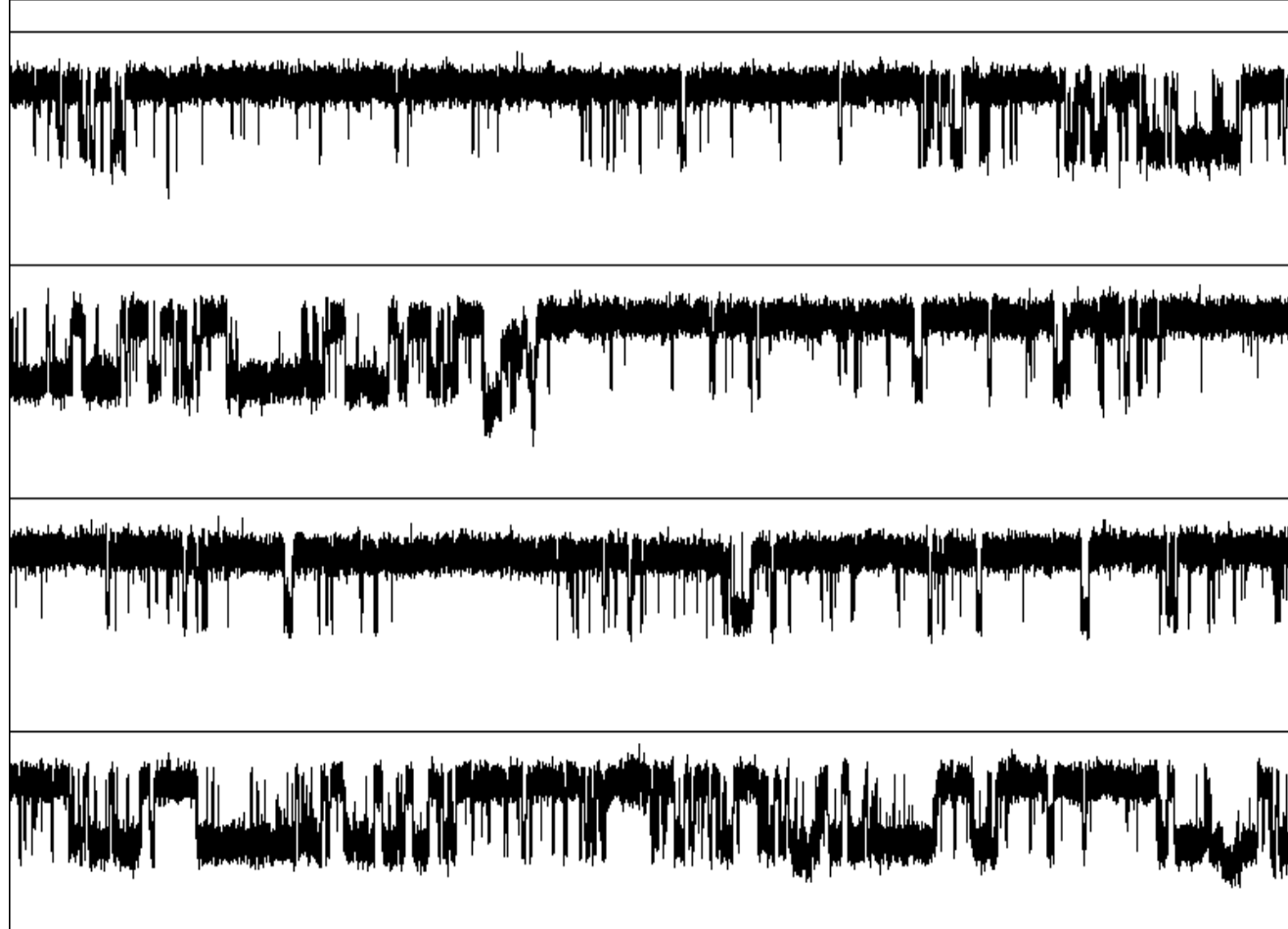


C:\temp\FREDGL~1.LDT

Drawing 0 - 399999, 100000 points/trace

Sampling frequency: 20 kHz

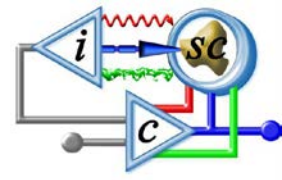
Low pass filter: 2 kHz



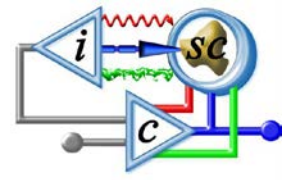
QuB(C) 2000-01 University at Buffalo, 10/16/2002 12:29:59 PM

Courtesy of A. Auerbach, SUNY-Buffalo

- Typical single-channel recordings of conductance fluctuations of a single gluR0 channel
- GluR0 in outside-out patch from HEK cells, zero  $\text{Ca}^{2+}$ , symmetric  $\text{K}^+$  saline, -60mV.



- Single ion channels might be useful as biosensors
  - Can be specific to a particular ligand
  - Can be natural or genetically engineered
  - Can be controlled by voltage or chemistry
  - Might allow DNA sequencing of bioagents
  - Best used in a high resistance lipid membrane
    - Can simulate expected local environment
    - Can allow normal function
    - Demonstrated single-channel sensitivity
    - Gigaohm seal reduces noise

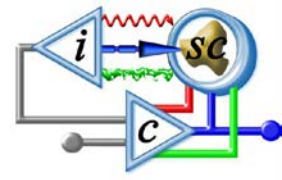


# The Appeal

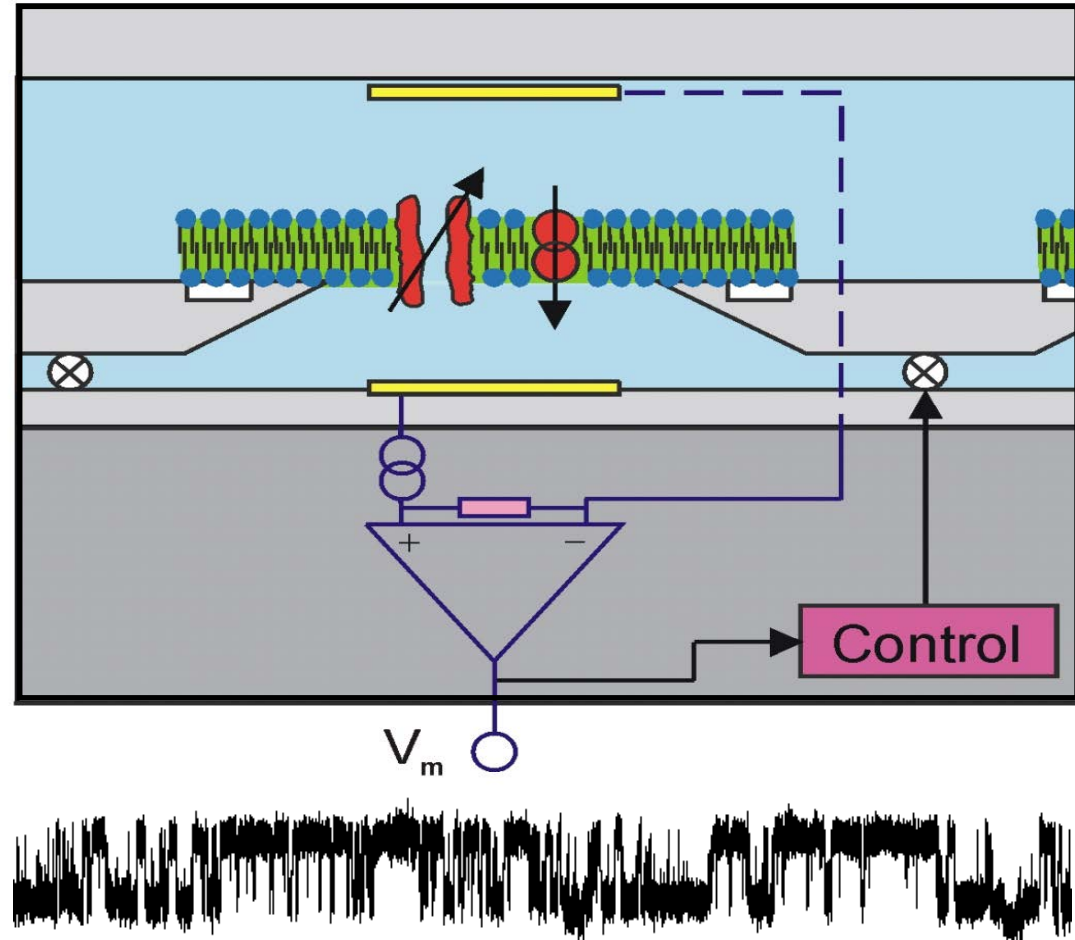
- In many cases, voltage- and ligand-gated channels and their signaling cascades exhibit a sensitivity, specificity, and efficiency of optimal physical devices such as photomultipliers
- Rhodopsin molecules and cyclic GMP-gated channels of retinal rods form single-photon detectors.
- Ion channels are high-gain amplifiers that transduce a small electrical, mechanical, or chemical signal into a large response
- A single ion channel can detect a single molecule.
- What are the limitations?



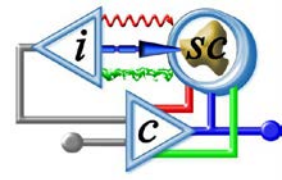
# How do you make an ion-channel biosensor?



- Make two small silicon bottles
- Connect with a small hole
- Cover the hole with a lipid membrane
- Put channel in the membrane
- Put different test solutions in one chamber
- Measure the current through the channel



# What is the gain of a ligand-gated ion channel?

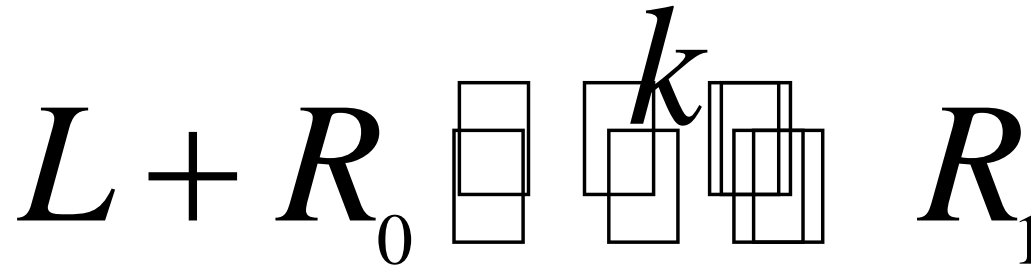
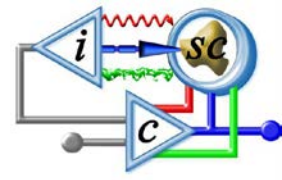


- Gain  $\equiv$  the number of ions that pass through the channel for one bound ligand

$$I = g \times V \quad Q = I \times t \quad N = \frac{Q}{z \times e} \quad N = \frac{g \times V \times t_{bound}}{z \times e}$$

- $1 \text{ ms} < t_{bound} < 10 \text{ ms}$
- $10^4 < \text{flux} < 10^7 \text{ ions per second}$
- $10 < \text{Gain} < 10^5$ .
- Large channels like gluR0 in normal  $[K^+]$  pass about  $10^7 \text{ ions/s}$  at a 100 mV driving force. In higher  $[K^+]$  or  $V_m$  they will pass more ions. The open time occurs in bursts that typically last for **one second**. **For these channels, the "gain", i.e., the integrated ion flux/ligand binding, is  $>10^7$**

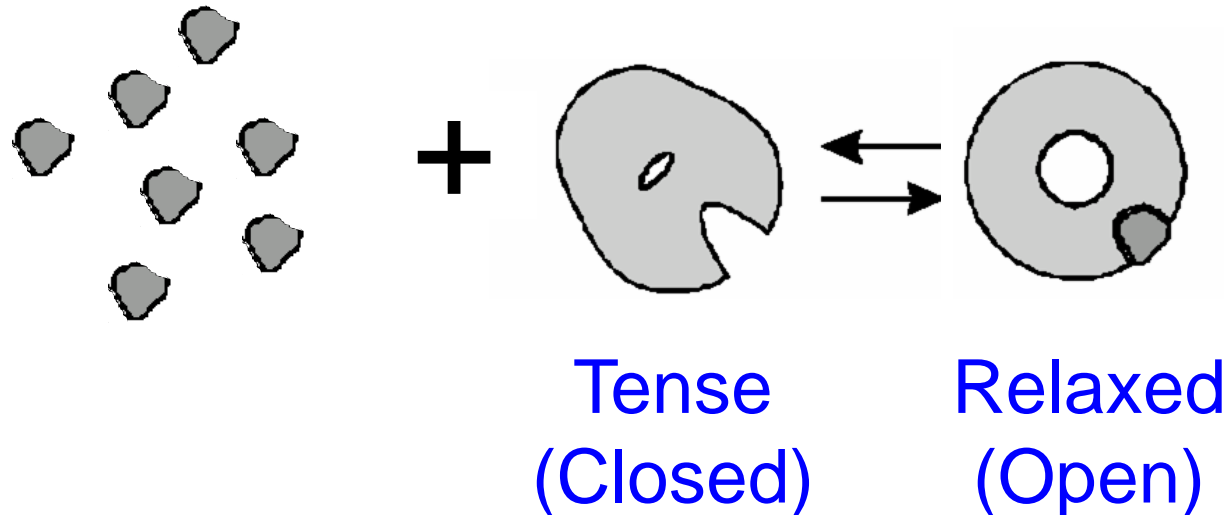
# Does it matter that the binding reaction is bimolecular?



Free Ligand

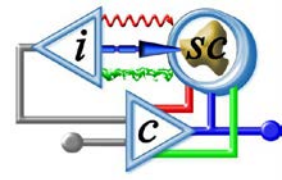
Unbound

Bound

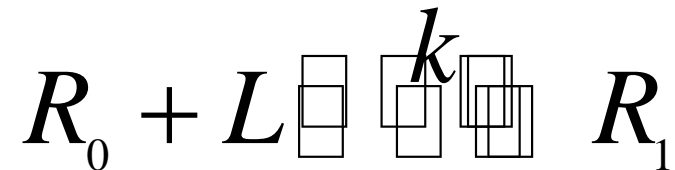


“Allosteric Mechanisms in the Activation of Ligand-Gated Channels,”  
 Meyer B. Jackson, University of Wisconsin - Madison  
<http://www.biophysics.org/btol/img/Jackson.M.pdf>

# Ligand binding is a bimolecular reaction



- Unbound Receptor plus Ligand gives Bound Receptor



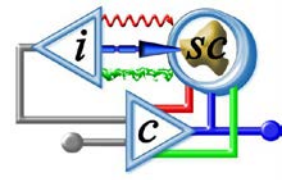
- The product of [Ligand] and [Unbound Receptor] are important.

$$\frac{[R_1]}{[L][R_0]} = k$$

- There is an energy associated with binding

$$\Delta G = -RT \ln(k)$$

# What you need

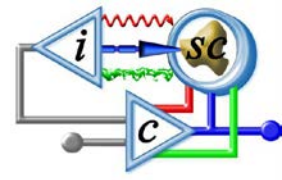


- You want the ligand to stick well

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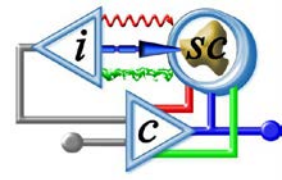


# What determines the reaction rate in a bimolecular reaction?

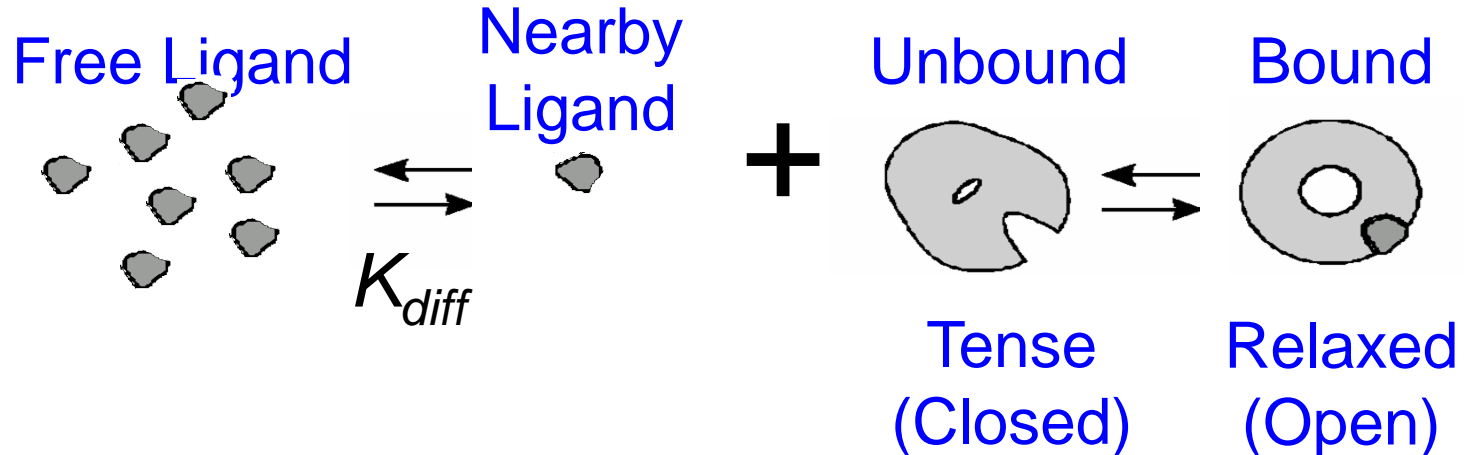


$$\frac{[R_1]}{[L][R_0]} = k$$

- The detector is a **SINGLE** receptor
- Assume one receptor in a (1 micron)<sup>3</sup> bottle
- **$[R] = 1.3 \times 10^{-24}$  mole/femtoliter = 1.3 nM**
- Binding rate =  $1/k$  events per second
- So what is the binding rate?

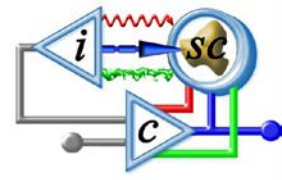


- For many reactions, the rate of reaction is determined by how long it takes a free ligand to diffuse into the vicinity of the receptor
- $K_{diff} \approx 10^8 \text{ M}^{-1}\text{sec}^{-1}$  so  $K_{on} \approx K_{diff} [L]$
- If  $[L] = 1 \text{ nM}$ ,  $1/K_{on} \approx 0.1 \text{ binding event/second}$

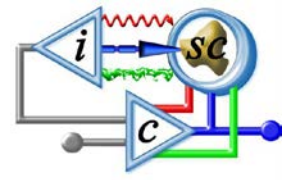


1 mM	1 $\mu\text{m}$	1 nM
10 $\mu\text{s}$	10ms	10 s

# *VI*<sub>BRE</sub> What happens once the ligand is bound?



- For a good sensor, once the ligand is bound, it should stay bound long enough to get a good measure of the conductance.
- To measure a concentration, rather than simply the presence of a single molecule, the sensor has to be reused, and hence the ligand must first unbind (dissociate).
- Suppose 10 seconds to measure and 100 seconds to recover.  $K_{off} > 0.01$ . Can detect  $[L] \approx 1$  nM
- 10 events will require 1000 sec or 17 minutes

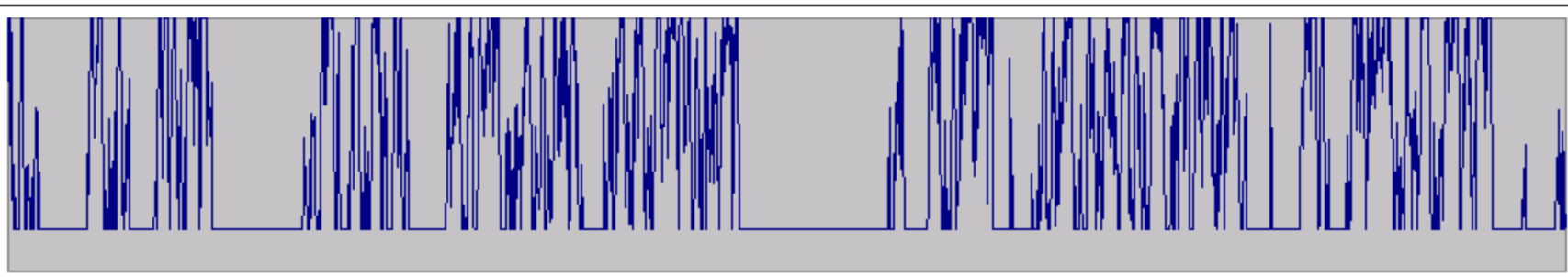


# What you need

- You want the ligand to stick well
- You don't want the ligand to stick so well that you can't reuse the channel
- You have to measure for a long time to get enough data to get a concentration

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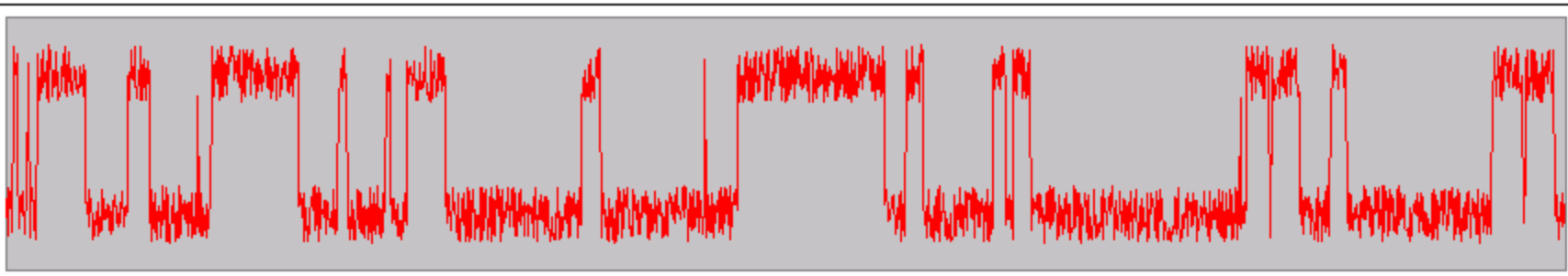
## Ligand Position



Diffusing

Bound

## Channel Current



Relaxed

Tense

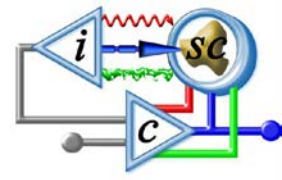
Average Channel Current 0.462

Binding Probability 0.950

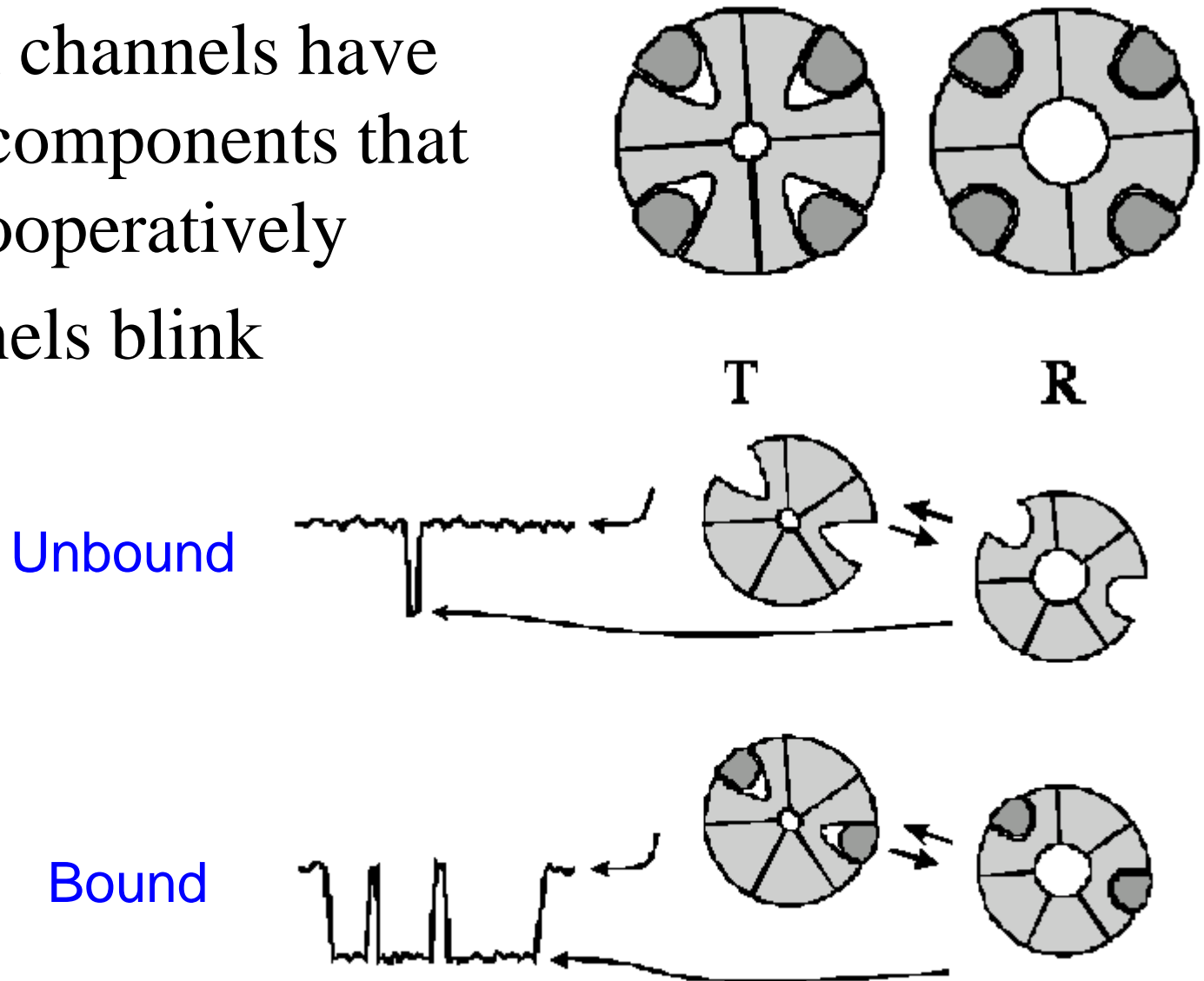
Unbinding Probability 0.020

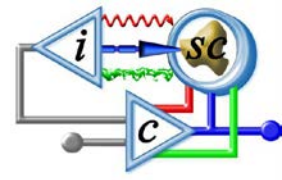


# What are the complications?



- Many ion channels have multiple components that behave cooperatively
- Ion channels blink

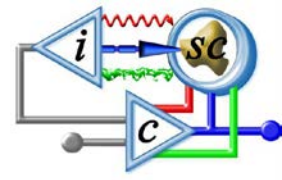




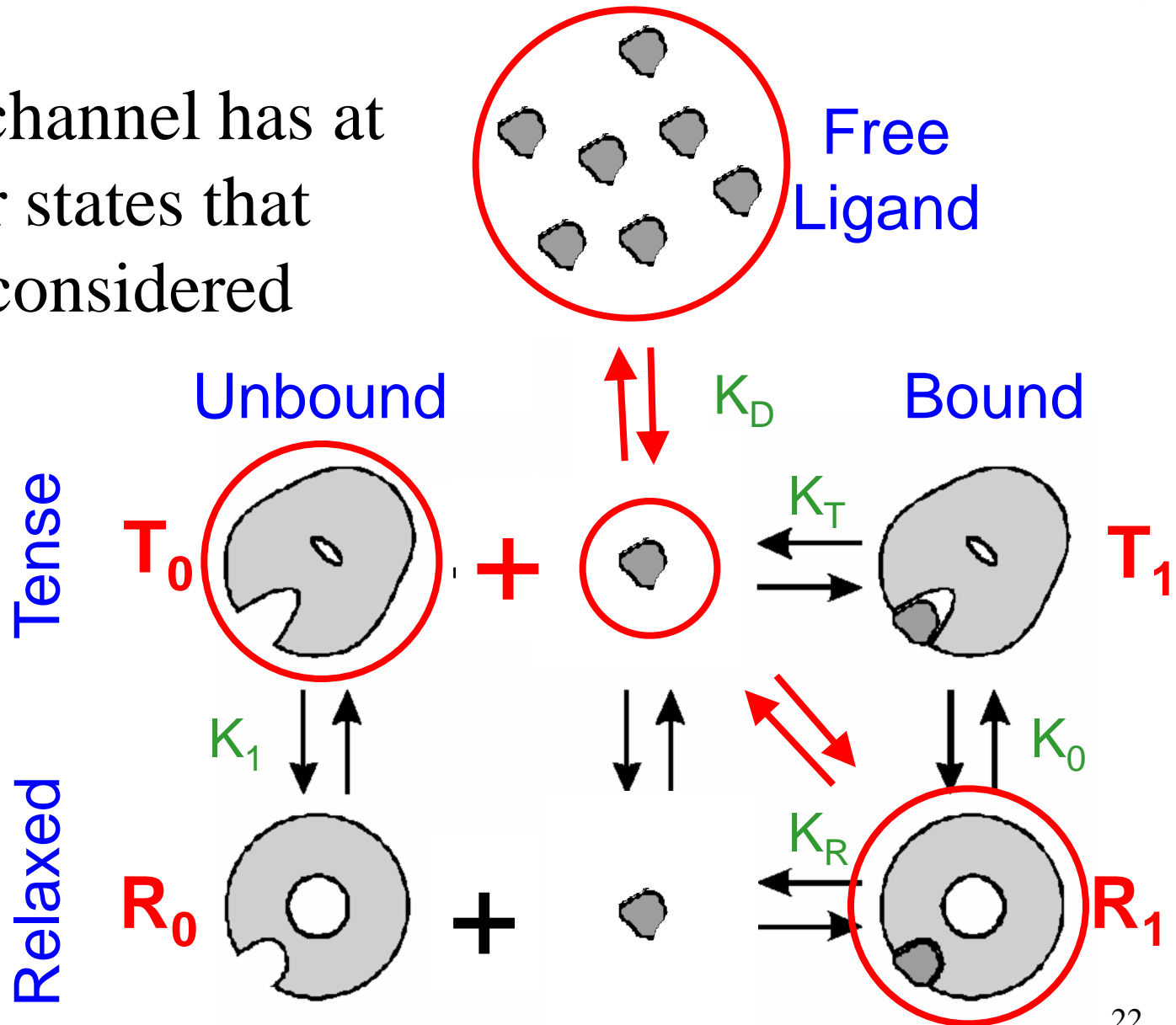
## What you need

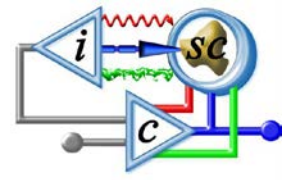
- You want the ligand to stick well
- You don't want the ligand to stick so well that you can't reuse the channel
- You have to measure for a long time to get enough data to get a concentration
- You want the ligand to stick well enough to minimize blinking

# A more detailed view



- The ion channel has at least four states that must be considered

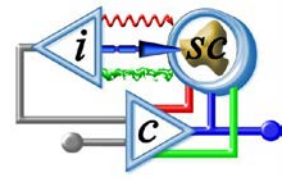




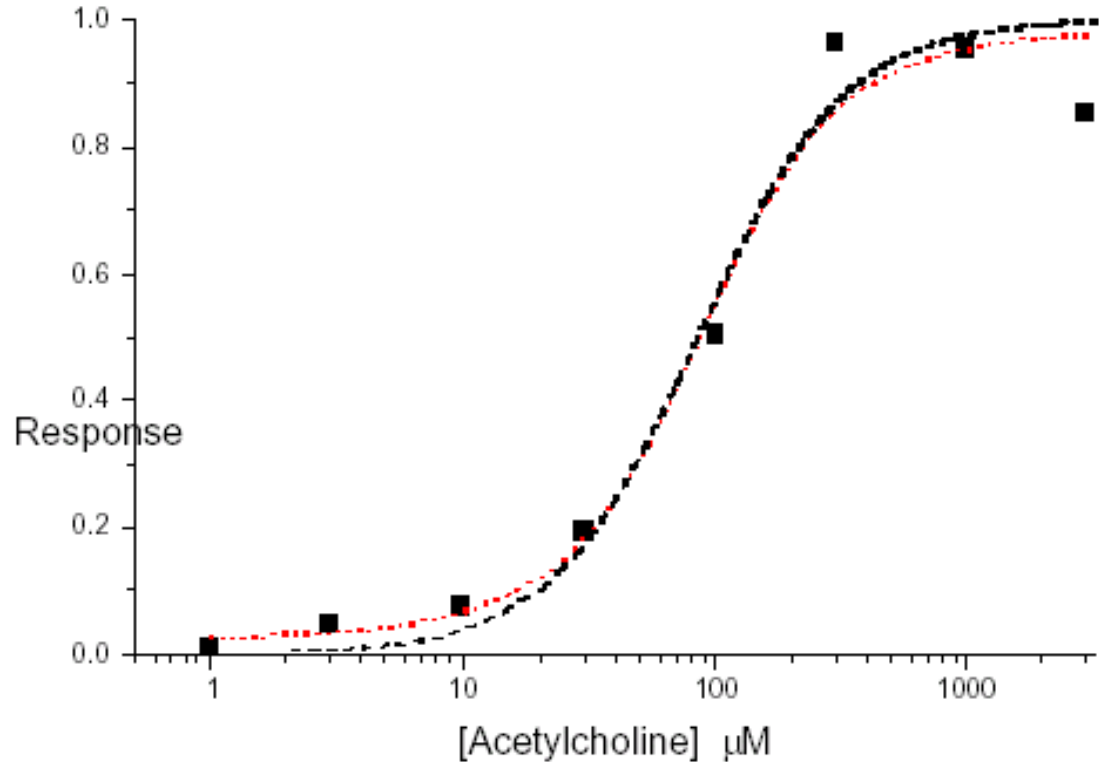
## What you need

- You want the ligand to stick well
- You don't want the ligand to stick so well that you can't reuse the channel
- You have to measure for a long time to get enough data to get a concentration
- You want the ligand to stick well enough to minimize blinking
- You need to measure a long time to get the channel kinetics

# Other problems

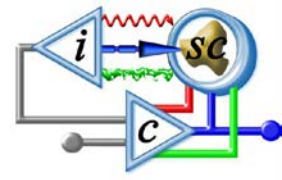


- Binding is not a binary event
- Binding is not perfectly specific
- As we said, many channels have multiple binding sites and cooperative binding

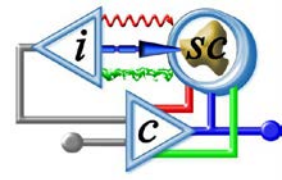




# Solutions to get faster response and bigger signals



- A single ion channel is infinitely sensitive if you wait infinitely long, but you then couldn't measure the concentration...
- Put the ligand molecule in a  $(0.1 \text{ } \mu\text{m})^3$  box to get  $[\text{L}] = 1 \text{ } \mu\text{M}$ . Detection straightforward thereafter
- Put multiple channels in the patch ( $\sim 500$ )
  - Increase the current
  - Increase the probability of getting a binding event
  - Loose information about channel binding dynamics
- Use a massively parallel array of ion channels



- Single channels have a very high internal gain as detectors where binding of one molecule can result in the transport of  $> 10^7$  ions.
- A single-channel chemical detector is not a single molecule detector: it runs on a bimolecular reaction with  $[R][L]$ .
- Single molecule sensors take time to respond that is dependent upon concentration in a diffusion-limited manner.
- To detect concentration, channel detectors must make repeated cycles of binding and unbinding since concentration is inferred from the time between binding events.
- While channels can be engineered to improve selectivity and responsiveness, diffusion places limits on the maximum speed of response.
- The use of channels as detectors requires the ability to distinguish different compounds in mixtures of different concentrations. This requires large parallel arrays.